Bottom-Up Approach to Synthetic Cells

Reinhard Lipowsky MPI of Colloids and Interfaces, Potsdam-Golm

- From Matter to Life
- Three Basic Modules:
  - Compartments, Motors, Assemblers
- New Platform for Bottom-Up Assembly
- Future Challenges

# From Molecules to Cells

#### **Transition Zone**

Aqueous Solution			••••		
Monomers	Polymers	Biocolloids Biomodules		Prokaryotes Organelles	Eukaryotes

... Matter

# Universal Architecture of All Cells

All present-day cells contain the same type of molecules and molecular processes -> All cells arose from a common ancestor



# Origin of Life



# Rough Time Schedule

- 4.6 Ga Solid earth
- 4.0 Ga Large lunar craters
- 3.8 Ga Sedimentary rock -> Water
- 3.5 Ga First microfossils resembling cyanobacteria = procaryotes?
- 1.8 Ga First microfossils resembling red algae = eukaryotes?
- ?.? Ga Invention of meiosis and sex
- 0.6 Ga Multicellular organisms



# Prebiotic Synthesis of Amino Acids

#### • Miller and Urey Experiment (1953)



Aminosäure	Murchison- Meteorit	künstliche Ursuppe
Glycin		
Alanin		
α-Amino-n-Buttersäure		
α-Aminoisobuttersäure		• •
Valin		• •
Norvalin		
Isovalin	• •	• •
Prolin		•
Picolinsäure	•	•
Asparaginsäure		
Glutaminsäure		• •
β-Alanin	• •	• •
β-Amino-n-Buttersäure	•	•
β-Aminoisobuttersäure	•	•
γ-Aminobuttersäure	•	• •
Sarkosin	• •	
n-Ethylglycin	• •	
n-Methylalanin	• •	

# Puzzle: Proteins + Nucleic Acids

• "Entanglement" of proteins and nucleic acids :





DNA contains blueprints of proteins Proteins perform replication, transcription of DNA

- Egg and hen problem, Eigen's paradoxon
- Common ancestor arose from prebiotic evolution: RNA world, iron-sulphur world, clay world ... ?

# Approaching the Transition Zone

Bottom-Up -> <- Top-Down



# Bottom-Up versus Top-Down

Bottom-Up: Synthetic Cells

- Develop important building blocks or modules
- Assemble these modules into larger structures
- Integrate more and more modules ...

#### Top-Down: Minimal Cells

- Start with relatively simple cells
- Eliminate more and more components
- Problem: many remaining genes with unknown functions

### Three Basic Modules







- Membrane compartments, fluid architecture
- Molecular motors, free energy transduction

• Molecular assembly, ribosomes, protein synthesis

#### Membrane Fluidity

• Fluid membranes, i.e., fast lateral diffusion:

Diffusion constant ~  $\mu m^2/s$ 

- Lateral diffusion => Compositional responses, demixing, domain formation ...
- Flexibility => Morphological responses, budding, tubulation, ...
   Direct evidence for fluidity



#### lipid swapping ~ ns





40 μm

### Multiresponsive Behavior

- Giant unilamellar vesicles (GUVs), tens of micrometers
- Remodelling in response to various perturbations:



Nanotubes from polymer adsorption, tube width ~ 100 nm Formation of intramembrane domains, 2D phase separation Small buds from protein adsorption, bud size ~ μm Remodelling by adhering or partially wetting droplets

#### Buds and Nanotubes

Liu et al, ACS Nano (2016)

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- Lipid mixture of DOPC, DPPC, cholesterol
- Membranes labeled by fluorescent dyes
- Liquid-disordered (red) and liquid-ordered phase (green)



- Asymmetric environment, different PEG concentrations
- Deflation: Bud and tube formation without external forces
- Tubes can be necklace-like or cylindrical

## Multi-Compartments from Curv Elasticity

• Buds and necklaces, uniform membranes: • Buds and necklaces, multi-domain membranes:

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- Each shape = single membrane with membrane necks
- Each shape is stable for large parameter regime
- Key parameters: membrane area, vesicle volume, spontaneous (or preferred) curvature

#### Spontaneous Tension

RL, Faraday Discuss. (2013) Bhatia et al (under review)

- Spont curvature *m* generates spon tension  $\sigma = 2 \kappa m^2$
- Micropipette aspiration of tubulated vesicle:



Initial aspiration up to hemispherical tongue,

then vesicle starts to flow like a liquid droplet

• Vesicle behaves as liquid droplet with interfacial tension equal to spontaneous tension  $\sigma$ 

## **ESCRT-Induced Budding**

Avalos Padilla et al, unpublished

• Sequential addition of three ESCRT proteins to GUVs:



• Interpretation: Domain-induced budding

### **Protein-rich Droplets**

- Brangwynne ... Hyman, Science (2009)
- Membrane-less organelles that behave like liquid droplets
- Enriched in intrinsically disordered proteins (IDPs)
- Example for IDP: RNA-binding protein FUS
- Interaction of FUS-droplets with membranes, two subsequent wetting transitions:





dewetting for high salt

partial wetting for intermediate salt

complete wetting for low salt 17







- Membrane compartments, fluid architecture
- Molecular motors, free energy transduction

• Molecular assembly, ribosomes, protein synthesis

### **Biomolecular Machines**



• Intro: Stepping motors



• Structural remodelling: Actin filaments



• Transport: Motor teams



• Information processing: Ribosomes 19

### Multiscale Aspects of Mol Motors

• ATP hydrolysis ~ 1 nm



• Mechanical steps ~ 10 nm





 Cargo transport by motor teams ~ 100 μm •Traffic of many motors/cargos and phase transitions 20

# Nanoscale: Mechano-Enzymes

- Motor action based on ATP hydrolysis
- Motor = ATPase with several catalytic domains
   M = # catalytic domains ≤ # ATP binding sites
- Examples:

Kinesin:M = 2Myosin V:M = 2Dynein: $M = 2 - 4 \le 8$ 



F1 ATPase: M = 3 < 6GroEl : M = 7 < 14



# Mesoscale: Thermodynamics

• Motor molecule coupled to several reservoirs:



- Isothermal motor activity at fixed temperature T
- Chemical energy change  $\Delta \mu = \mu(ATP) \mu(ADP) \mu(P)$
- Mechanical work  $W_{me} = \ell F$  during spatial displacement  $\ell$

# State Space of Motor

RL et al: J. Stat. Phys 135 (2009)

- Motor states *i* and *j*
- Transition lij> from *i* to *j* with rate  $\omega_{ij}$



- Energy change during lij> arising from chemical potential difference  $\Delta \mu_{ij}$  and mechanical work  $W_{ij}$ :
  - $U_j U_i = \Delta \mu_{ij} Q_{ij} W_{ij}$  (first law of TD)
- Free energy change from constrained equilibrium:

$$H_{j} - H_{i} = \Delta \mu_{ij} - W_{ij} - k_{B} T \ln (\omega_{ij} / \omega_{ji})$$

• Entropy change from thermodynamic relation:

$$S_j - S_i = k_B \ln (\omega_{ij} / \omega_{ji}) - Q_{ij} / T = \Phi_{ij} - Q_{ij} / T$$
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## **Cyclic Balance Conditions**

- Summation of transitions along a complete directed cycle  $C_v^{d}$ , all state functions cancel
- Released heat:  $Q(C_v^d) = \Sigma Q_{ij} = \Delta \mu(C_v^d) W(C_v^d)$
- Produced entropy I:  $T \Phi(C_v^d) = \Sigma T \Phi_{ij} = Q(C_v^d)$
- Produced entropy II:  $T \Phi(C_v^d) = k_B T \ln(\Xi_v^d)$

with 
$$\Xi_{v}^{d} = \prod_{ij>}^{v,d} (\omega_{ij} \neq \omega_{ji})$$

$$k_{B}T \ln(\Xi_{v}^{d}) = \mu(C_{v}^{d}) - W(C_{v}^{d}) = Q(C_{v}^{d})$$

Relation between kinetics and thermodynamics, must be fulfilled for thermodynamic consistency 24 • Balance condition for each directed cycle  $C_v^{d}$ :

$$k_{\rm B}T \ln(\Xi_{\rm v}^{\rm d}) = \mu(C_{\rm v}^{\rm d}) - W(C_{\rm v}^{\rm d})$$

Classification of cycles:

- Detailed balance:  $\mu(C_v^d) = 0$  and  $W(C_v^d) = 0$
- Mech nonequilibrium:  $\mu(C_v^d) = 0$  and  $W(C_v^d) \neq 0$
- Chem nonequilibium:  $\mu(C_v^d) \neq 0$  and  $W(C_v^d) = 0$
- Chemomech coupling:  $\mu(C_v^d) \neq 0$  and  $W(C_v^d) \neq 0$

### Cargo Transport by Motor Teams

• Transport by N identical motors

Klumpp and RL, PNAS (2005)

- Transport by two antagonistic motor teams, Stochastic tug-of-war

M. Müller et al, PNAS (2008)



• Elastic linkers between motors and cargo

Berger et al, *PRL* (2012) Ucar, RL, *Soft Matter* (2017)





# **Concentration Gradients from Motors**

M. Müller et al, J. Phys. CM 17 (2005)

• Half open tube:

left boundary open, reservoir of motors = 'cell body'
right boundary closed = 'Synapse'

• (+) Motors (kinesins) moving to the right



• (-) Motors (dyneins) moving to the left



Concentration gradient created by motors







- Membrane compartments, fluid architecture
- Molecular motors, free energy transduction

• Molecular assembly, ribosomes, protein synthesis

# Protein Synthesis by Ribosomes

- Ribosomes are assemblies of rRNA and r-proteins
- Complex and hierarchical assembly process in vivo
- In vitro assembly from rRNA and r-proteins without additional components



- No assembler for ribosomes
- Protein synthesis requires many molecular players:



#### Ribosome + mRNA + tRNAs



TC = ternary complex = tRNA + EF-Tu + GTP

EF-Tu = most abundant protein

- Ribosome steps along codons of mRNA (purple -> green) consuming one ternary complex at each codon
- Elongation cycle during one step:

Decoding of codon by binding/accommodation of tRNA Elongation of growing peptide chain by one amino acid Translocation of mRNA together with two tRNAs

# Single Elongation Cycle



• Complexity of decoding:

61 sense codons and 43 elongator tRNA species (E. coli)

# Codon-tRNA Relationships

- red/purple = non-cognate released after initial binding
- yellow = near-cognate decoding => wrong amino acid
- green = cognate decoding => correct amino acid
  - ,Ocean' of non-cognates with some near-cognates and a few cognates



### Single Elongation Cycle - Refined

Rudorf, Thommen, Rodnina, RL, PLoS Comp Biol (2014)

• Possible binding of cognate/near-cognate/non-cognate tRNAs:



• Competition between cognate, near-cognate, and non-cognate tRNAs

#### Markov Process

• Map cartoon of multistep process onto Markov chain:



• Individual transitions:

initial binding, recognition, initial selection, GTP hydrolysis, phosphate release, proof reading, full accommodation

- All transition rates  $\omega_{ij}$  have been measured in vitro
- Some rates identical for both cognates and near-cognates <sup>34</sup>

#### From In-Vitro to In-Vivo Rates Rudorf, Thommen, Rodnina, RL, PLoS Comp Biol (2014) • Scale factors $\omega_{ii}^*/\omega_{ii}$ • Single barrier shifts $\Delta_{ij} = \ln(\omega_{ij}^*/\omega_{ij})$ 0.5 Single Barrier Shift A<sub>ij</sub> [k<sub>B</sub> T] Scale Factor $\omega^*_{ m ij}/\omega_{ m ij}$ -0.5 -1.0-1.53th 3th 3th 3th 3th 3th 3th 3th 3th 300

• Three in-vivo rates (purple) are significantly increased: rejection rate  $\omega_{76}$  for near cognates dissociation rate  $\omega_{off}$  after initial binding recognition rate  $\omega_{rec}$  for cognates and near-cognates 35

## Assembly of Protein Complexes

#### Shieh ... Kramer, Bukau, Science (2015)

- Co-translational assembly: *in vivo* synthesis of two proteins, assembly during translation, luxA binds to emerging luxB
- Protein synthesis on a chip: different DNA compartments for different proteins, control of spatial separation between different compartments





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Karzbrun ... Bar-Ziv, Science (2014)

## New Platform for Bottom-Up Assembly

Weiss ... Spatz, Nature Materials (Nov. 2017) #

- Collaboration within MaxSynBio
- Project leader: Joachim Spatz, Heidelberg
- Collaboration between 4 MPIs and 3 Universities
- Water-in-oil emulsion droplets
- Generated by microfluidics, stabilized by surfactant

#### # List of coauthors:

MarianWeiss, Johannes Patrick Frohnmayer, Lucia Theresa Benk, Barbara Haller, Jan-Willi Janiesch, Thomas Heitkamp, Michael Börsch, Rafael B. Lira, Rumiana Dimova, Reinhard Lipowsky, Eberhard Bodenschatz, Jean-Christophe Baret, Tanja Vidakovic-Koch, Kai Sundmacher, Ilia Platzman and Joachim P. Spatz

# GUVs within W/O Emulsion Droplets

- Emulsion w/o droplet stabilized by surfactant
- Pico-Injection of small vesicles
- Pico-Injection of Mg<sup>++</sup>
- Adhesion of vesicles to surfactant layer
- Rupture of vesicles
- Fusion of fragments

- Image: Surger of the surger
- => Formation of a GUV supported by surfactant layer
- Release of encaged GUV from droplet

### Release of GUVs from Droplets

Weiss ... Spatz, Nature Materials (Nov. 2017)



• General platform for subsequent bottom-up assembly

## Sequential Bottom-Up Assembly



- Pico-injection of membrane and cytoskeletal proteins
- Incorporation of functional ATP Synthase

### Perspectives and Challenges

- Further steps of sequential assembly: Compartments + ATP Synthase + motors + ...
- Importance of ionic conditions
- ,Broken hierarchy' of biolevels
- Wanted and unwanted interactions
- Evolution via selection (failures)
- Evolution as a learning process
- Natural cells after 10<sup>8</sup> years ?
- Synthetic cells after xx years ?
- Can science replace evolution?





#### Summary



Lipid bilay

- Membrane compartments, multiresponsive, many architectures
- Molecular motors, cargo transport and concentr gradients
- Protein synthesis, comparison of in vivo and in vitro
- Droplet-stabilized GUVs, new platform for sequential assembly





• Membranes

Rumiana Dimova Tom Robinson Jaime Agudo-C. Tripta Bhatia Yunuen Avalos Padillo Jan Steinkühler • Motors + Ribosomes Stefan Klumpp Sophia Rudorf Mehmet Ucar Stefanie Foerste Nadin Haase Simon Christ Collaborations
Marina Rodnina
Joachim Spatz
Tony Hyman
Titus Franzmann
Günther Kramer
Roy Bar-Ziv