

# Forces in Biosystems

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- Theory and Biosystems
- Synthetic Biology, Bottom-Up
- Forces Generated by Membranes
- Outlook on Mol Machines
- Perspectives and Challenges

# Basic Aspects of Theory

Criteria for a ‘good’ theory:

- Consistent with fundamental laws
- Consistent with experimental/simulation data
- Helpful to understand experimental/simulation data
- General relations between experimental observables
- Predictive power: interesting, nontrivial predictions

Pleasures of theory:

- Quantitative predictions confirmed by experiment
- New insight into underlying mechanisms
- Aesthetic appeal

# Some Quotes

„Nothing is more practical than a good theory“

Immanuel Kant

„As simple as possible but not simpler“

Albert Einstein

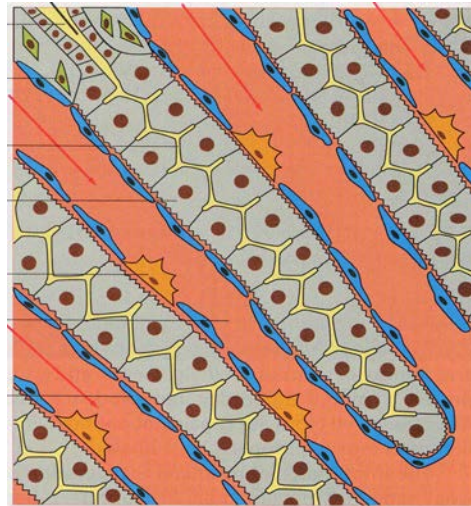
„A good theory is like a good joke: it is short and the last line is quite unexpected“

H. L. Friedman

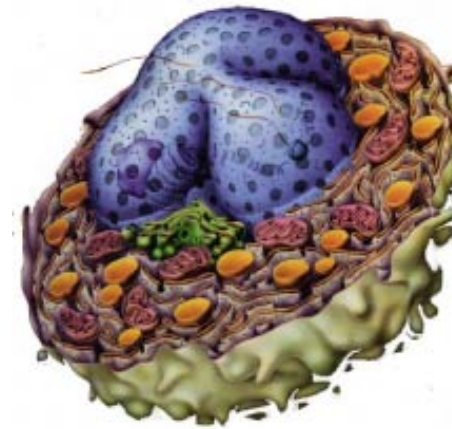
# Biosystems: Top-Down



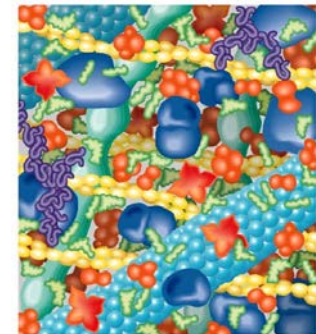
Human body  
[m]



Tissues  
[mm]

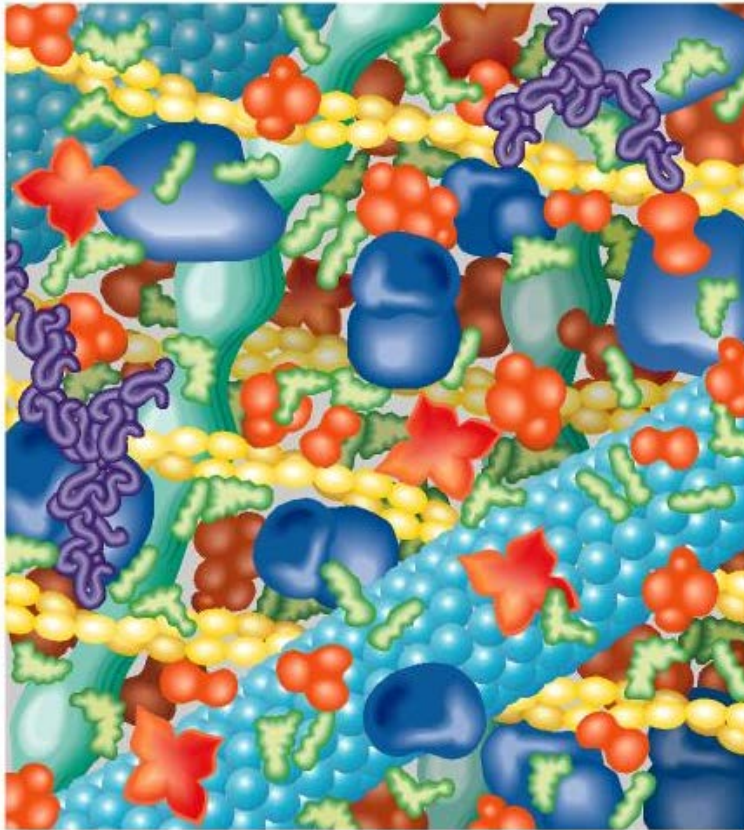


Cells  
[ $\mu\text{m}$ ]



Molecular  
Assemblies  
[nm]

# Universal Nanostructures



25 nm  
Bsp: Filament

- Water and ions
- Small molecules (monomers) form **macromolecules** such as
  - Proteins
  - RNA, DNA
  - Lipids
  - Polysaccharides
- Macromolecules form **molecular assemblies** such as
  - Ribosomes
  - Filaments
  - Membranes ....

# ‘Selforganization’

- Assembly of building blocks "by themselves"
- Instructions from local environment

## (1) Selforganization via molecular interactions

Structure formation **close to** equilibrium

Examples: Protein folding, Binding, Adhesion, ...

## (2) Selforganization via free energy transduction

Chemomechanical coupling **far from** equilibrium

Examples: Molecular motors, Filament assembly, ...

- **Biological** systems are difficult to understand because they exhibit ‘entanglement’ of (1) and (2)
- ‘Disentangle’ via **biomimetic** model systems

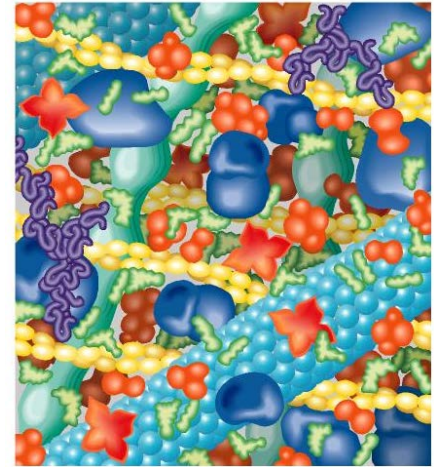
# Hidden Dimensions

- Molecular and nanoscopic building blocks are

small + flexible + mobile

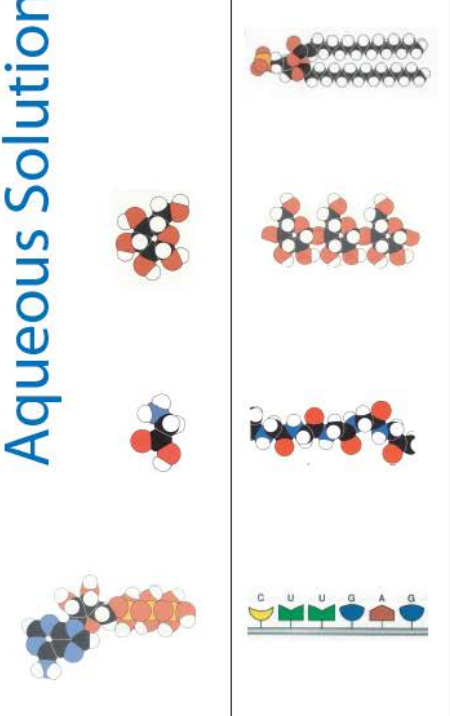
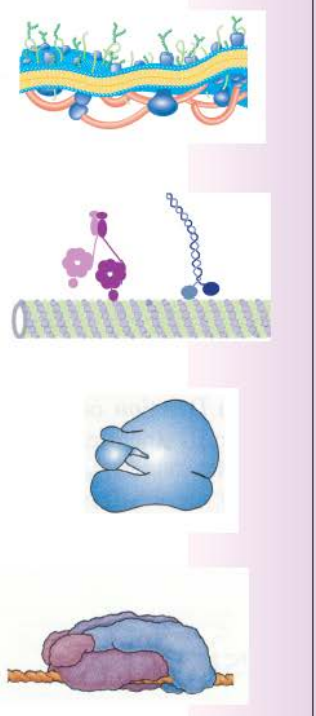
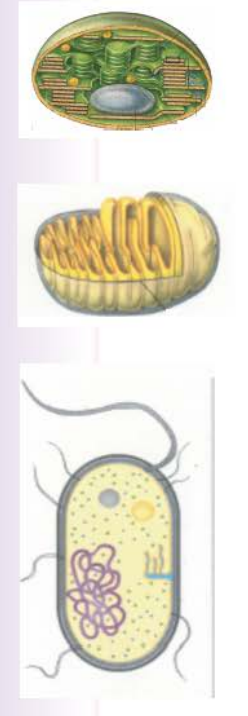
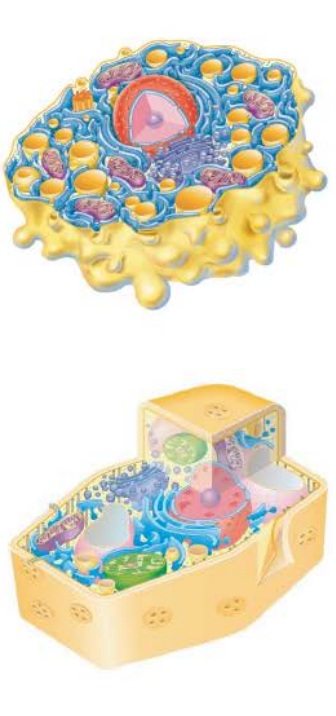
Length scales: nm up to  $\mu\text{m}$

Time scales: ns up to min



- No single experimental method can cover all scales
- High spatial resolution but low temporal resolution,  
No experimental nanoscope
- High spatio-temporal resolution by simulations
- Theory: Unification of experimental and simulation data

# Biosystems: Bottom-Up

Aqueous Solution			Transition Zone		
Monomers	Polymers	Biocolloids Biomodules	...	Prokaryotes Organelles	Eukaryotes

Matter

Life



# Two Routes into Transition Zone

## Bottom-Up: Synthetic Cells

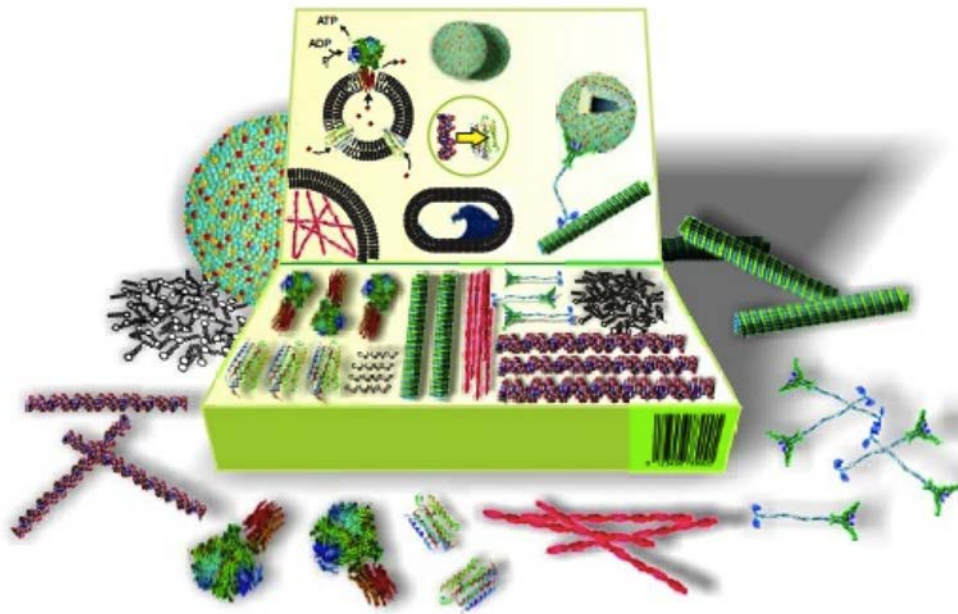
- Develop important building blocks or modules
- Assemble these modules into larger structures
- Integrate more and more modules ...

## Top-Down: Minimal Cells

- Start with relatively simple cells
- Eliminate more and more components
- Problem: many remaining genes with unknown functions

# Synthetic Biology, Bottom-UP

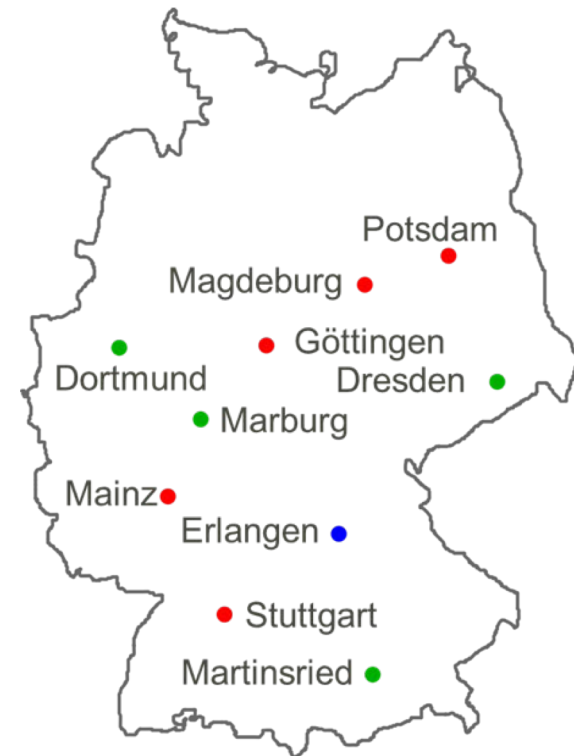
MaxSynBio Consortium:  
Create a toolbox of modules  
to build a synthetic cell



Motivation:

No understanding of minimal  
cell from top-down approach

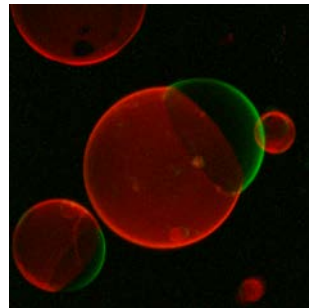
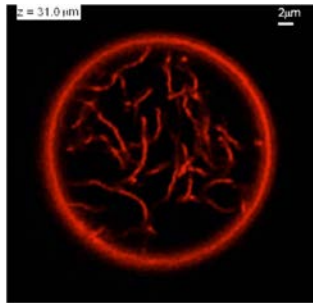
9 Max Planck Institutes:



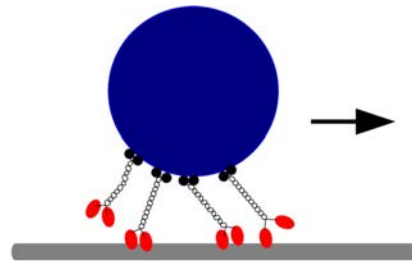
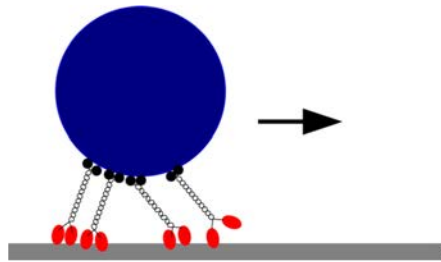
Speakers:

Petra Schwille,  
Kai Sundmacher

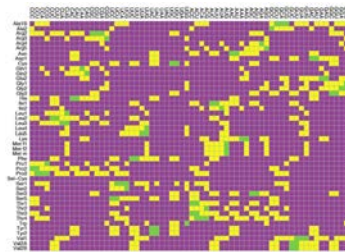
# Basic Modules for Synthetic Cells



- Membrane and vesicles, fluid compartments, remodeling



- Directed transport by molecular motors, free energy transduction

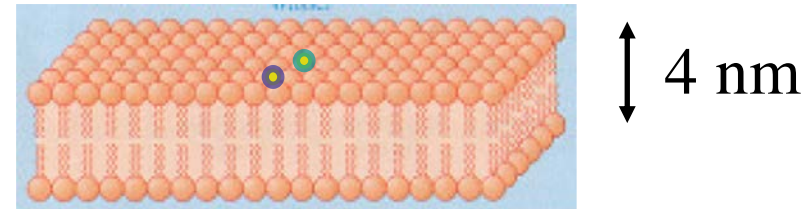


- Template-directed assembly, ribosomes, protein synthesis

# Biomembranes are Fluid Bilayers

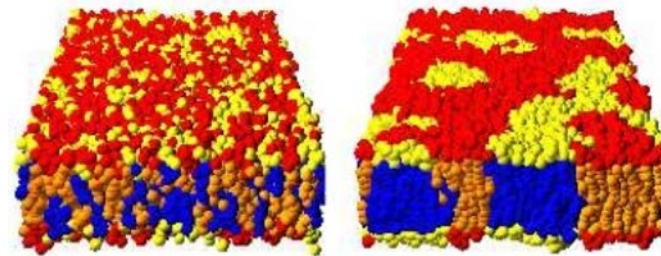
- **Fluid** membranes, i.e.,  
fast lateral diffusion:

Diffusion constant  $\sim \mu\text{m}^2/\text{s}$

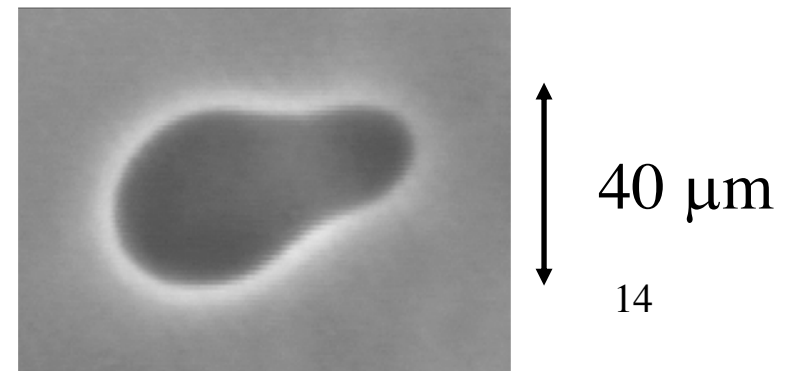


lipid swapping  $\sim \text{ns}$

- Lateral diffusion =>  
**Compositional responses,**  
demixing, domain formation ...

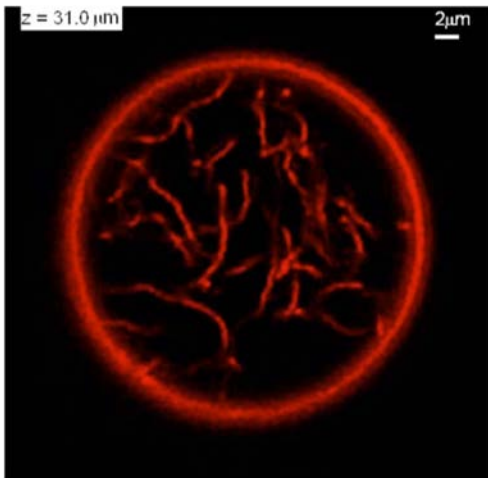


- Flexibility =>  
**Morphological responses,**  
budding, tubulation, ...  
Direct evidence for fluidity

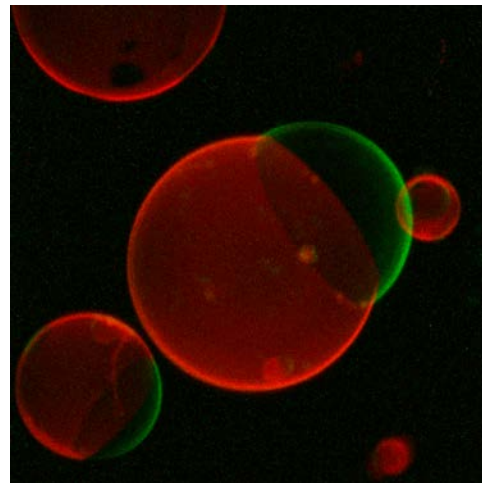


# Multiresponsive Behavior

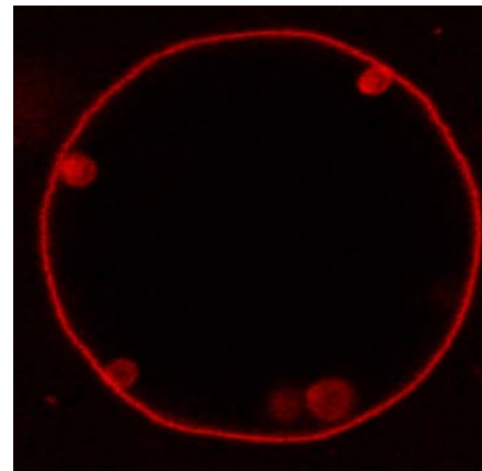
- Giant unilamellar vesicles (GUVs), tens of micrometers
- Remodelling in response to various perturbations:



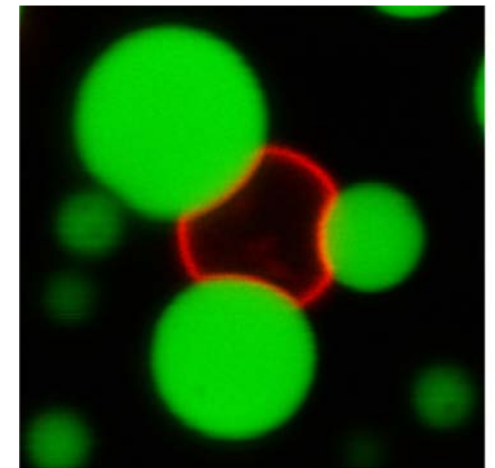
Nanotubes from polymer adsorption, tube width  $\sim 100$  nm



Formation of intra-membrane domains, 2D phase separation



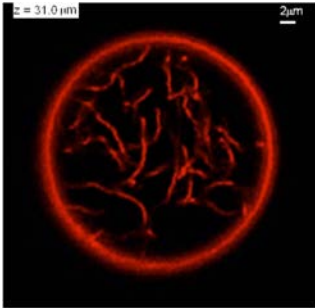
Small buds from adsorption of two ESCRT proteins



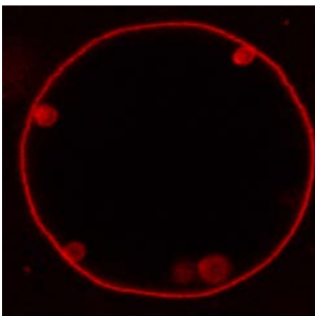
Shaping GUVs by membrane-less organelles, FUSb

- What are the **forces** that drive remodelling processes?

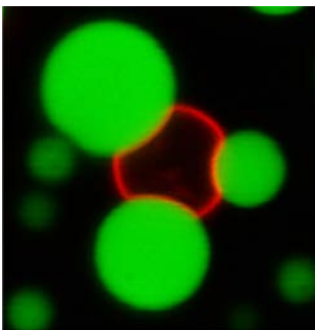
# Forces Generated by Membranes



- Spontaneous Tubulation:  
Spont curvature generates spont tension  
and constriction forces



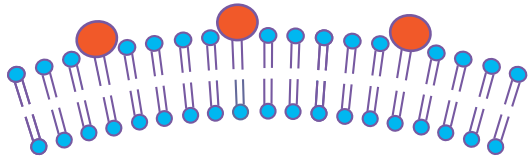
- ESCRT-induced budding and fission:  
Adhesion-induced constriction forces



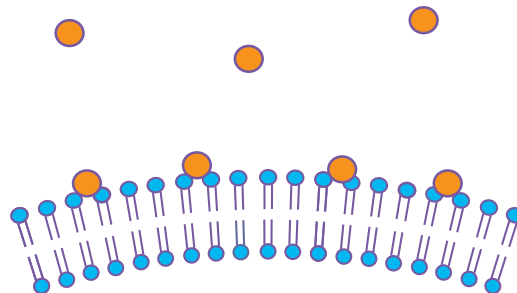
- Interactions with membrane-less organelles:  
Capillary forces and curvature generation

# Spontaneous = Preferred Curvature

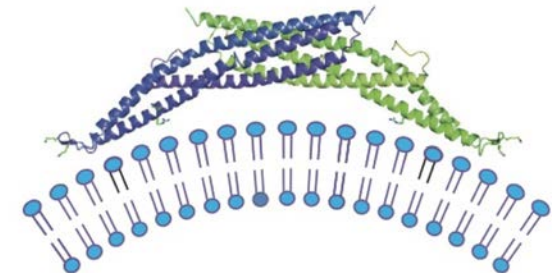
- Spontaneous or preferred curvature  $m$  describes bilayer asymmetry = asymmetry between two leaflets
- Different molecular mechanisms for bilayer asymmetry:



Asymmetric  
composition,  
e.g., ganglioside



Asymmetric  
adsorption of  
small molecules

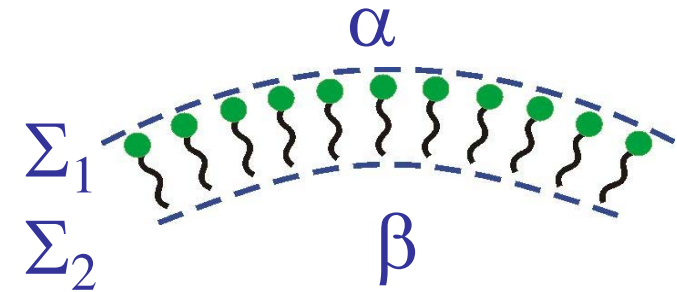


Asymmetric  
protein coats,  
e.g. BAR-domain

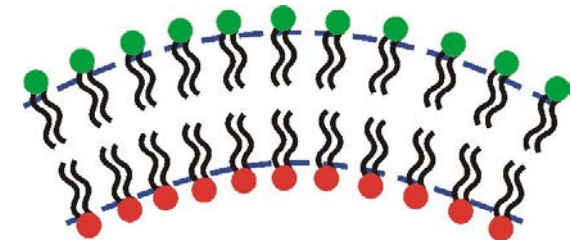
# Concept of Spontaneous Curvature

- W. D. Bancroft (1913)  
‘Theory of emulsification’
- F. C. Frank (1958)  
‘On the theory of liquid crystals’
- W. Helfrich (1973)  
‘Elastic properties of lipid bilayers’
- Variants of curvature models:

E. Evans, S. Svetina + B. Zeks, M. Wortis



splay term from  
symmetry arguments





# Curvature Elasticity

- Mean curvature  $M$  tries to adapt to spontaneous (or preferred) curvature  $m$

- Curvature or bending energy:

$$E_{cu} = \int dA 2 \kappa (M - m)^2$$

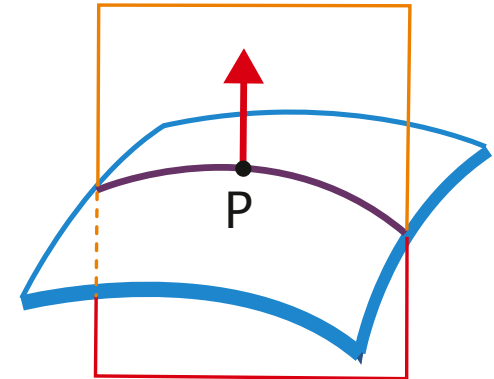
integral over membrane area  $A$

- 2nd fluid-elastic parameter: Bending rigidity  $\kappa$

Dimensions of energy,  $\kappa = 10^{-19} \text{ J} = 20 \text{ k}_B \text{ T}$

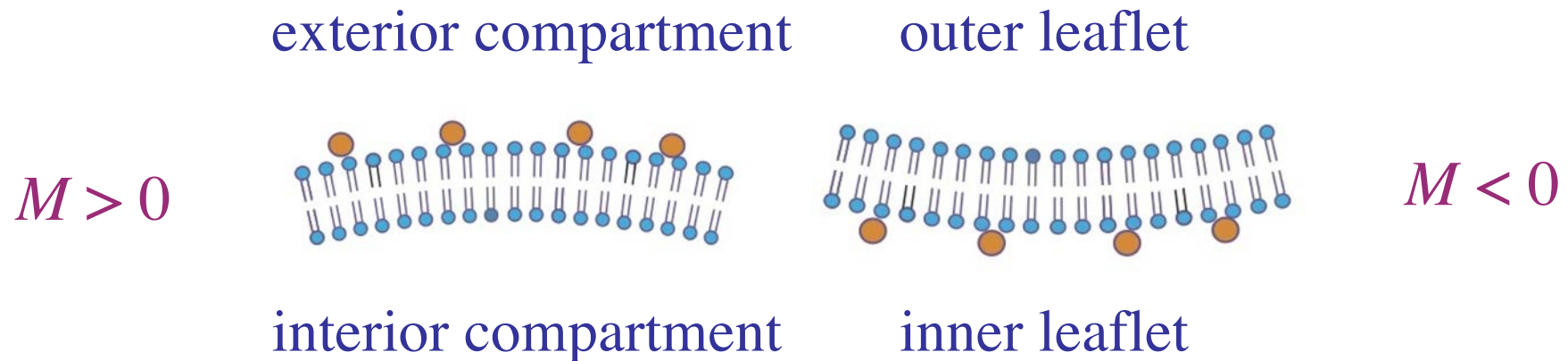
- Range of spontaneous curvatures  $m$

from  $1/(20 \text{ nm})$  to  $1/(20 \mu\text{m})$



# Sign of (Spontaneous) Curvature

- Mean curvature  $M$  and spontaneous curvature  $m$  can be positive or negative
- Sign defined with respect to interior/exterior compartments = with respect to inner/outer leaflet



Mean curvature  $M$  is positive (negative) if membrane bulges towards exterior (interior) compartment

# Shape Functional for Vesicles

- Vesicle has constant surface area  $A$  and fixed volume  $V$
- Shape functional:

$$F = E_{\text{cu}} + \Sigma A - \Delta P V$$

Mechanical tension

Pressure difference

- Consider  $\Sigma$  and  $\Delta P$  as Lagrange multipliers
- Minimization with respect to normal displacements of membrane, Euler Lagrange equation:

$$\Delta P = 2 \Sigma M - 2 \kappa \Delta_{LB} M - 4 \kappa [M - m] [M (M + m) - G]$$

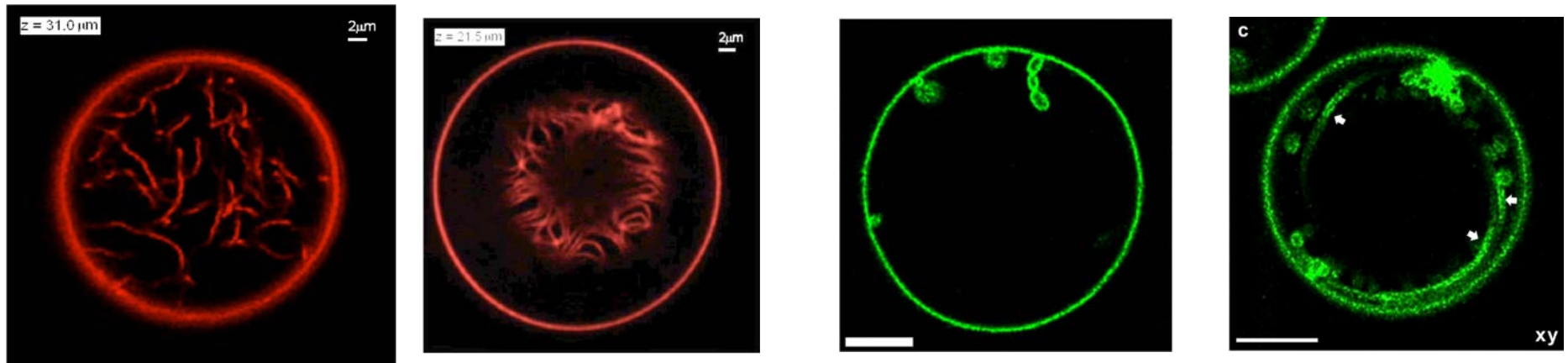
- Laplace equation plus  $\kappa$ -dependent terms
- Explicit solutions in reduced shape spaces

Ou-Yang, Helfrich,  
*Phys. Rev. A* (1989)

# Buds and Nanotubes

Liu et al, *ACS Nano* (2016)

- Lipid mixture of DOPC, DPPC, cholesterol
- Membranes labeled by fluorescent dyes
- Liquid-disordered (red) and liquid-ordered phase (green)

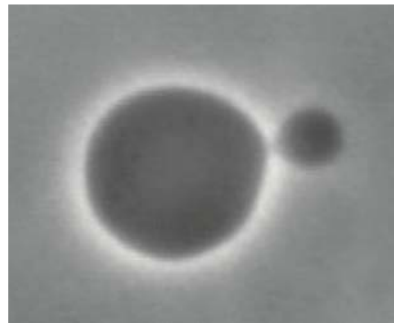
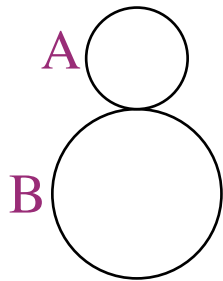


- Asymmetric environment, different PEG concentrations
- Deflation: Bud and tube formation **without** external forces
- Tubes can be necklace-like or cylindrical

# Membrane Necks

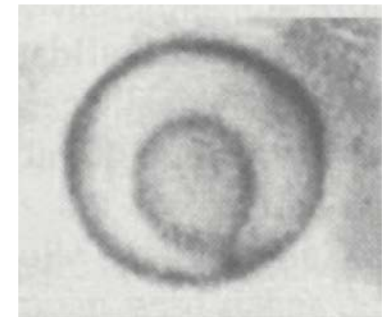
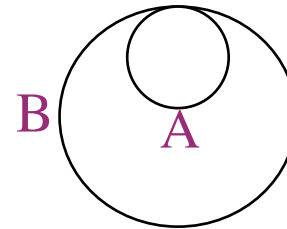
- For  $m \neq 0$ , curvature elasticity leads to spherical membrane segments connected by membrane necks

- Out-bud:



spont curv  $m > 0$

- In-bud:



spont curv  $m < 0$

- Closed neck is stable if:

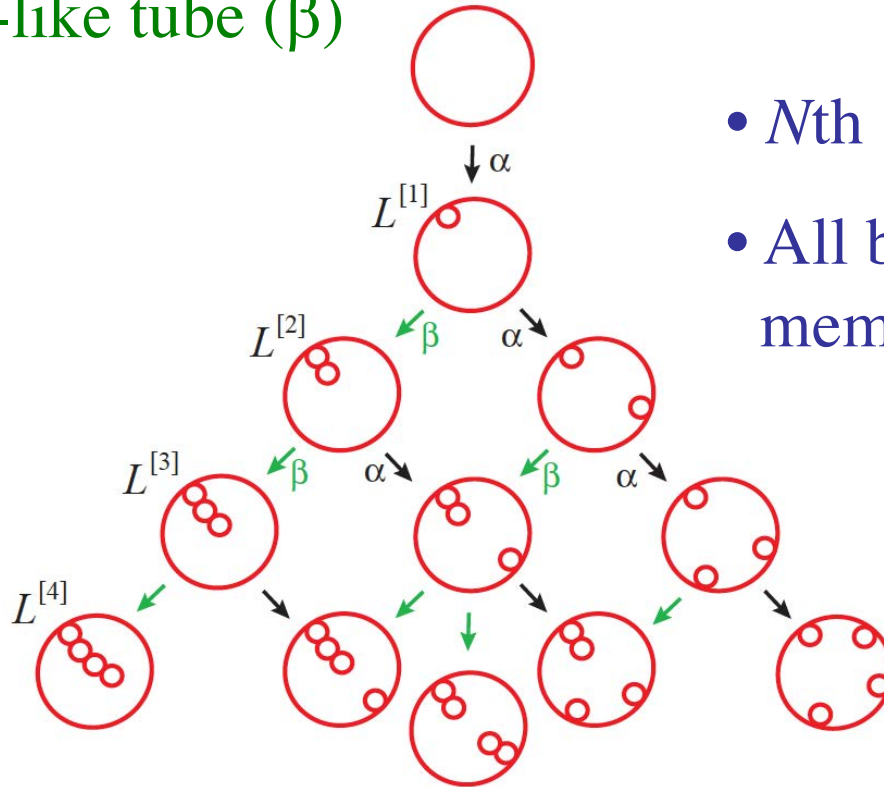
$$0 < M^A + M^B \leq 2m$$

$$2m \leq M^A + M^B < 0$$

- Relation between geometry and material parameter

# Nucleation and Growth of Tubes

- Vesicle membrane with large spont curv  $m$  Liu et al, *ACS Nano* (2016)
- Osmotic deflation of GUV in discrete steps
- At each step, nucleation of new bud ( $\alpha$ ) or extension of necklace-like tube ( $\beta$ )

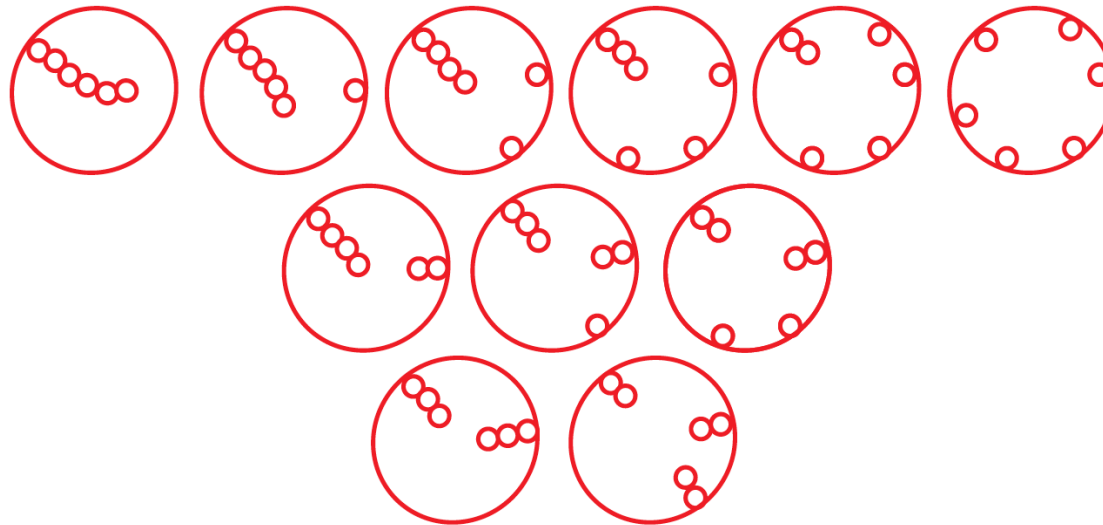


- $N$ th step leads to  $N$  in-beads
- All beads are connected by membrane necks (not visible)

=> Buds are nuclei for necklace-like tubes

# Morphological Complexity

- After 6th step, 11 morphologies with 6 beads:



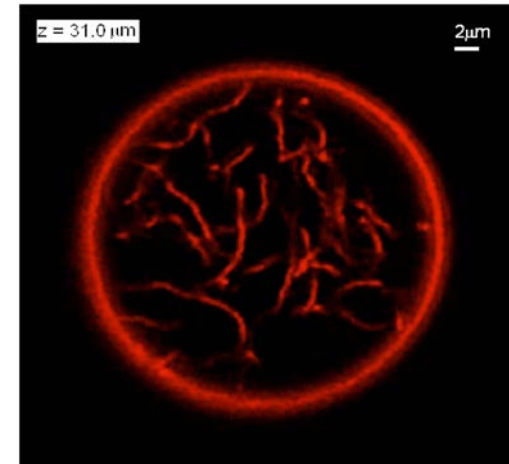
- All beads are connected by membrane necks
- All morphologies have the same area, volume, and curvature energy
- Rugged energy landscape contains 11 intersecting branches
- For large  $N$ , # of  $N$ -bead morphologies grows as  $\exp[c \sqrt{N}]$

# Spont Curvature Generates Tension

RL, Faraday Discuss. (2013)

- Tubulation leads to tense mother vesicle
- Total tension in Euler-Lagrange equation has two components:

$$\hat{\Sigma} = \Sigma + \sigma$$



Mechanical tension  $\Sigma$  stretches the membrane

Spontaneous tension  $\sigma = 2 \kappa m^2$  curves the membrane

- Presence of nanotubes implies dominance of spontaneous tension, mechanical tension can be ignored
- Example: Spont curvature  $\approx -1/(100 \text{ nm})$  implies

Spontaneous tension  $\sigma \approx 10^{-2} \text{ mN/m}$

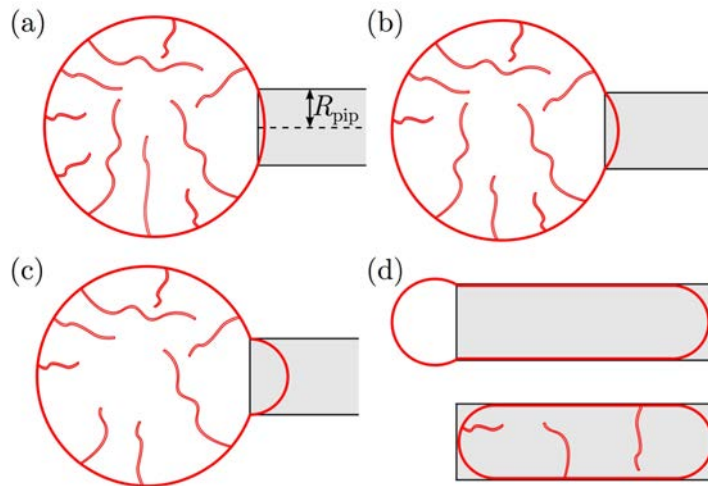
Mechanical tension  $\Sigma \approx 10^{-4} \text{ mN/m}$



# Spont Tension from Experiment

- Retraction of tubes by micropipettes:

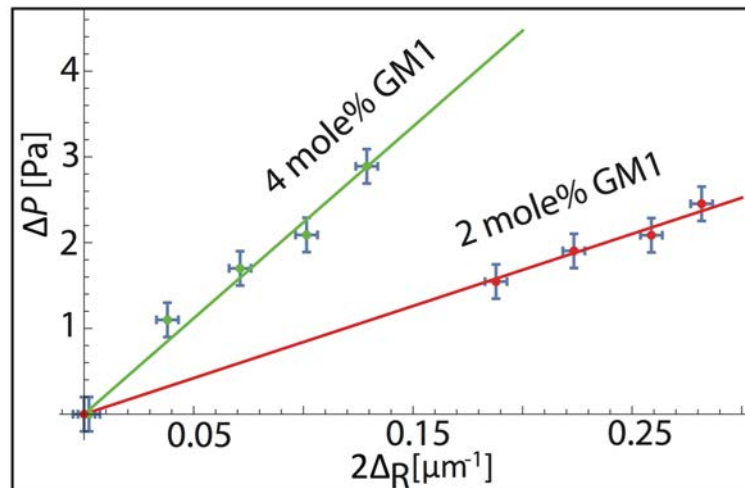
Bhatia et al, ACS Nano  
(under revision)



Initial aspiration up to hemispherical tongue

then vesicle starts to flow into micropipette, increased robustness !

Aspiration pressure

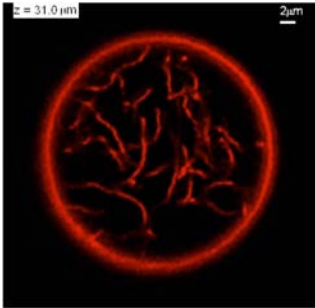


Initial aspiration:

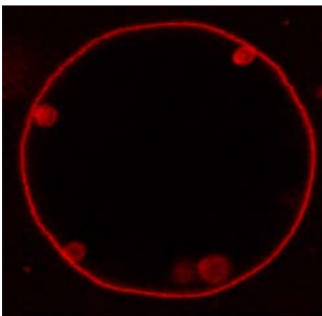
Aspiration pressure versus geometric quantity  $\Delta_R$

Slope = spontaneous tension  $\sigma$

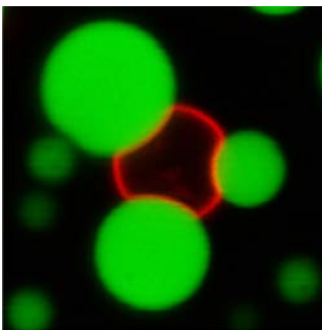
# Forces Generated by Membranes



- Spontaneous Tubulation:  
Spont curvature generates spont tension  
and constriction forces



- ESCRT-induced budding and fission:  
Adhesion-induced constriction forces



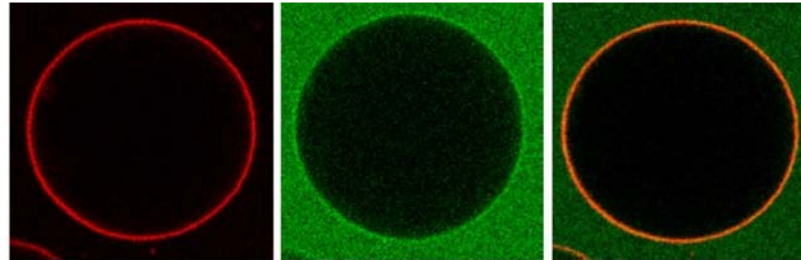
- Interactions with membrane-less organelles:  
Capillary forces and curvature generation

# Sequential ESCRT Addition

Avalos Padilla et al, *Frontiers Microbiology* (in press)

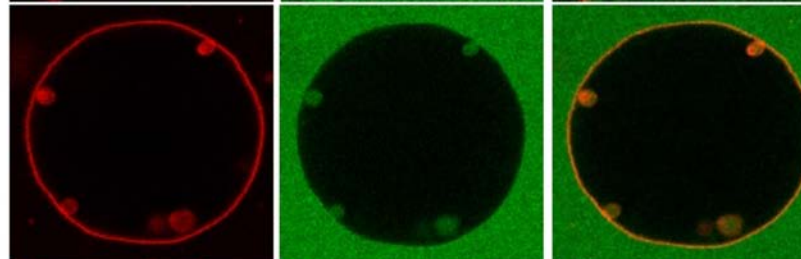
- Sequential addition of three ESCRT proteins to GUVs:

+ ESCRT 1  
Vps20



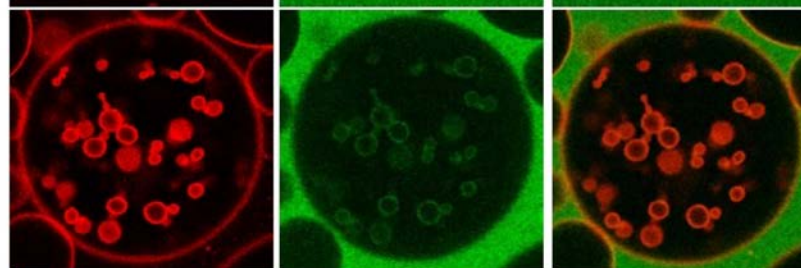
adsorption  
of Vps20

+ ESCRT 2  
Vps32



multiple  
buds + necks

+ ESCRT 3  
Vps24



fission of  
necks

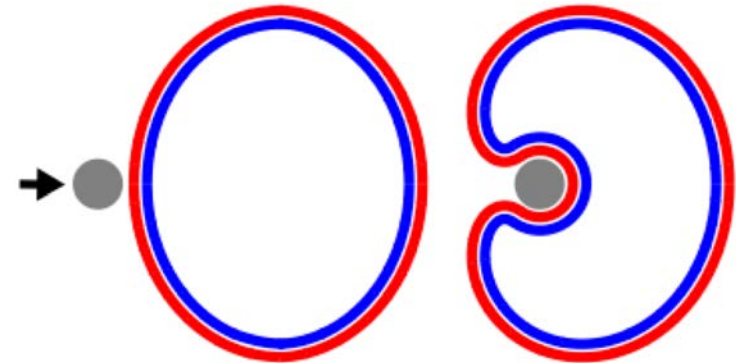
# Digression on Nanoparticles

Agudo-Canalejo, RL, *ACS Nano* (2015)  
*Nano Letters* (2015)



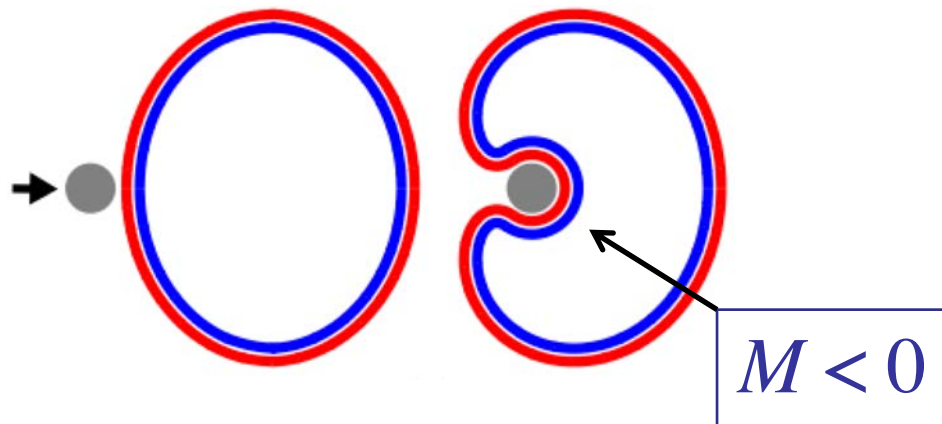
Jaime Agudo

- Nanoparticles interacting with membranes, vesicles and cells: biomedical imaging, drug delivery, nanotoxicity, virus infection ...
- Important control parameters:
  - Adhesive strength  $W \sim$  surface chemistry
  - Particle size  $R_{pa}$
  - Spontaneous curvature  $m$



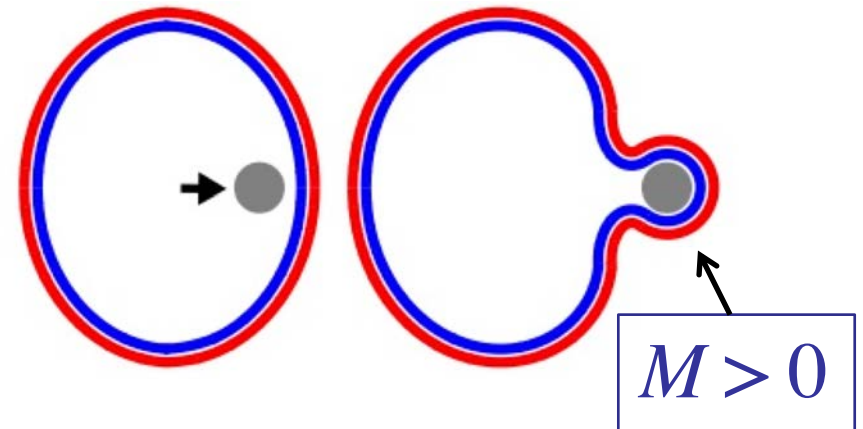
# Endo- and Exocytosis

- Endocytic engulfment:



- Particles originate from **exterior** compartment
- Negative curvature  $M$  of bound membrane segment
- Favored by  $m < 0$

- Exocytic engulfment:



- Particles originate from **interior** compartment
- Positive curvature  $M$  of bound membrane segment
- Favored by  $m > 0$

# Shape Functional with Adhesion

- Shape functional with adhesive term:

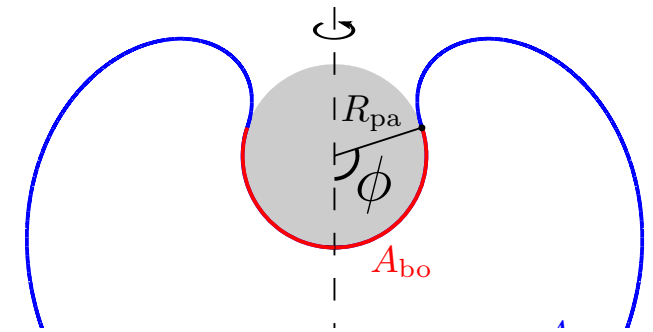
$$F = E_{cu} + \Sigma A - \Delta P V - W A_{bo}$$

Adhesion (free) energy

Adhesive strength  $W > 0$

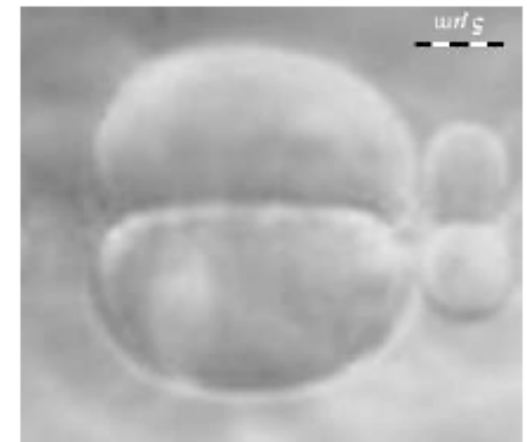
Area  $A_{bo}$  of bound membrane segment

Boundary bound / unbound segment



- Competition between adhesion and bending encoded in adhesion length  $R_W = (2\kappa/W)^{1/2}$
- Contact mean curvature  $M_{co}$  for membrane adhesion to planar surface:

$$M_{co} = 1/R_W = (W/2\kappa)^{1/2}$$



# Contact Mean Curvature at Particle

- Membrane adhering to particle
- Bound membrane segments (red) follows particle surface
- Contact line provides boundary condition for unbound segment (blue)
- Principial curvatures along contact line:

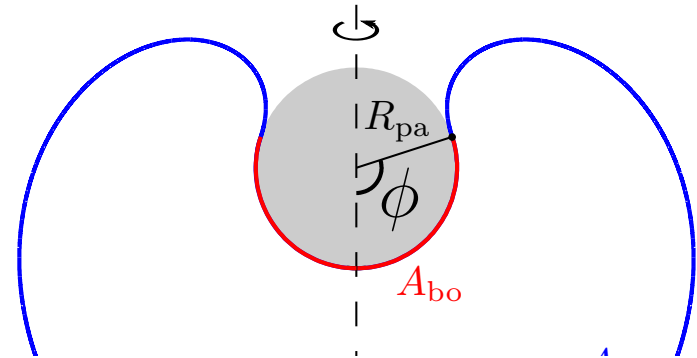
$$C_1 = (2W/\kappa)^{1/2} - 1/R_{pa} \quad (\text{along contour})$$

$$C_2 = -1/R_{pa} \quad (\text{perp to contour})$$

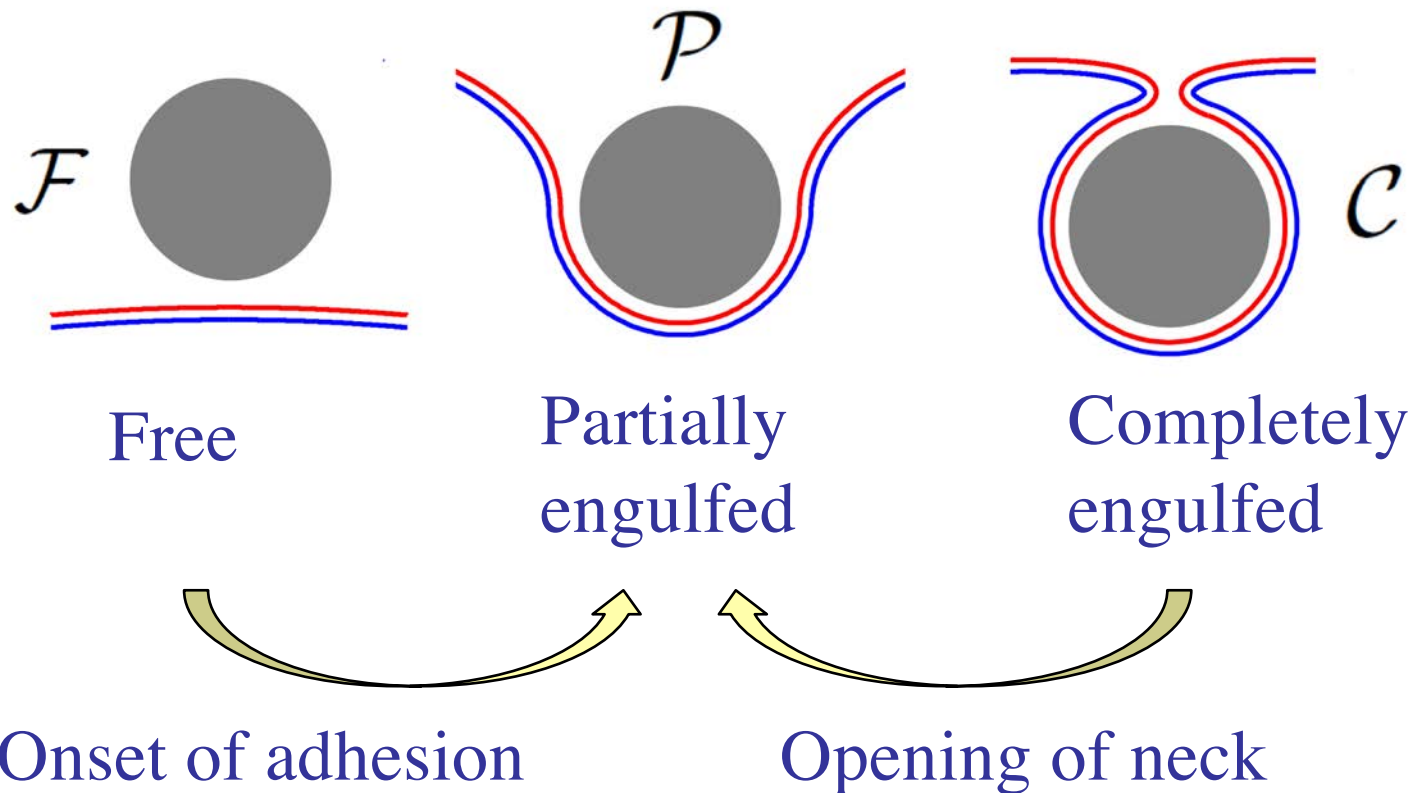
- Contact mean curvature:

$$M_{co} = 1/R_W - 1/R_{pa}$$

- Independent of spontaneous curvature  $m$  !



# (In)Stability of Particle States

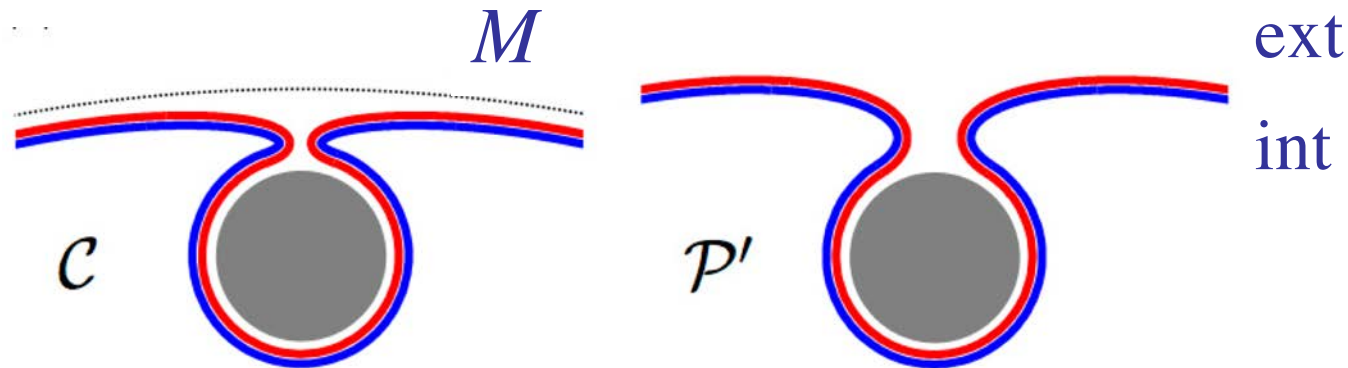


- Competition between adhesion and bending:

Adhesion length  $R_W = (2\kappa/|W|)^{1/2}$



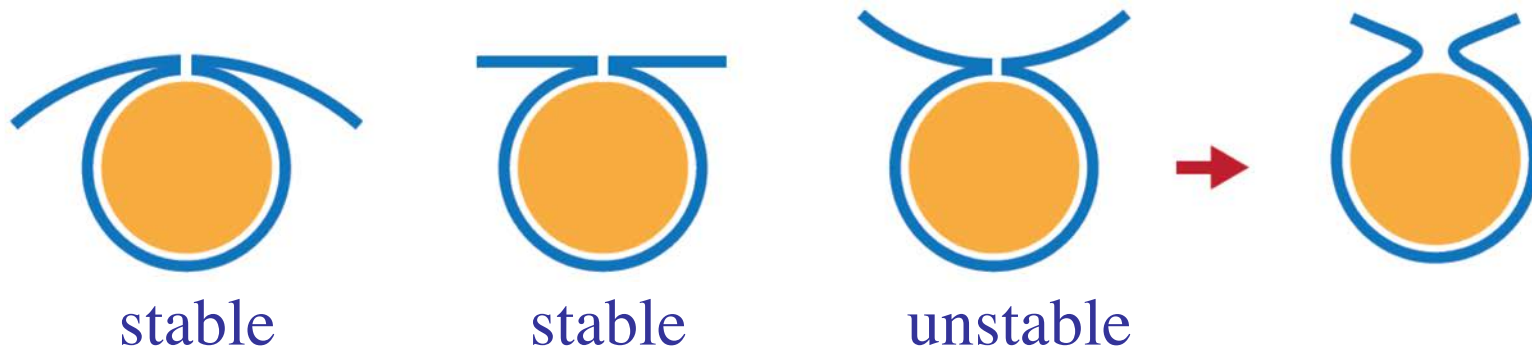
# Opening of Membrane Neck



- Closed neck is stable if mean curvature

$$M \geq 2m - M_{co} = 2m - 1/R_W + 1/R_{pa}$$

- Example: threshold value  $2m - M_{co} = 0$  :



# Clathrin-dependent Endocytosis

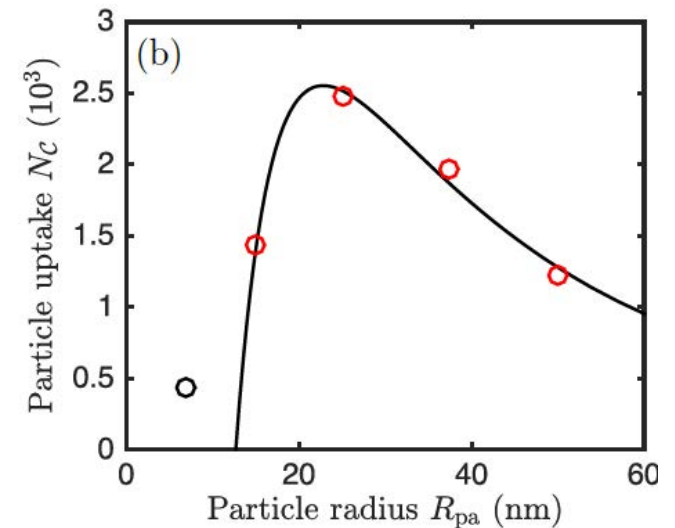
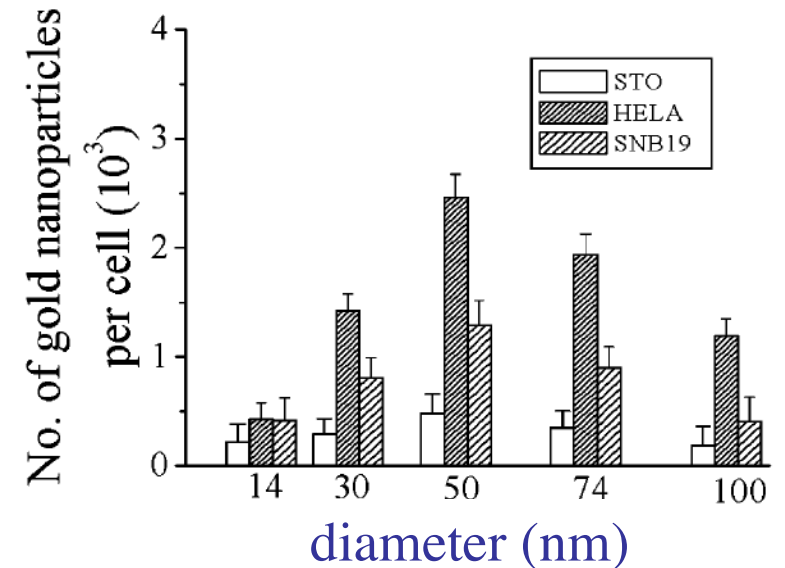
Chithrani et al, *Nano Letters* (2007)

- Uptake of gold nanoparticles by cells
- Particles bind to transferrin receptors
- Assembly of clathrin-coated vesicles

Non-monotonic size-dependence !

- Cell membrane with two types of segments, bound and unbound
- Bound segment contains protein coat with spont curv  $m_{b0} = -1/(40 \text{ nm})$
- Good agreement with exp data:

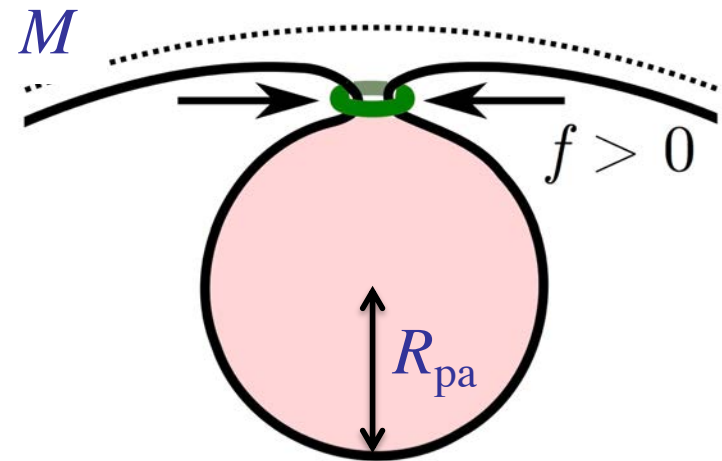
Agudo-Canalejo, RL: *ACS Nano* (2015)



# Stability of Membrane Necks

Agudo-Canalejo, RL, *Soft Matter* (2016)

- Necks of in-buds stabilized by
  - constriction force  $f$
  - (negative) spontaneous curvature  $m$
  - substrate with adhesive strength  $W$



- General stability relation for  $m < 0$ :

$$M_{pa} = -1/R_{pa}$$

$$f(4\pi\kappa)^{-1} - 2m + (|W|/2\kappa)^{1/2} - 1/R_{pa} + M \geq 0$$

- Linear superposition of different mechanisms

# Effective Constriction Force

- General stability relation for  $m < 0$ :

$$f (4\pi\kappa)^{-1} - 2m + (|W|/2\kappa)^{1/2} - 1/R_{pa} + M \geq 0$$

- Effective constriction force

$$f_{\text{eff}} = f + 8\pi\kappa|m| + 2\pi(2\kappa|W|)^{1/2}$$

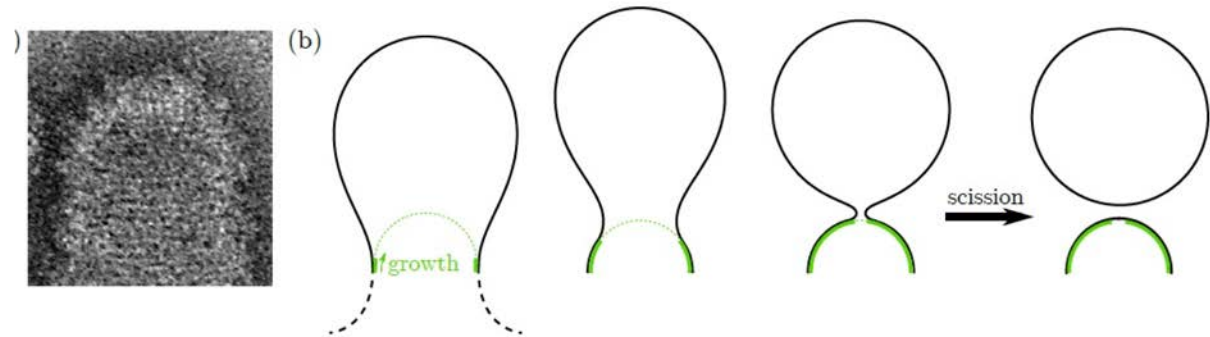
- Spontaneous curvature generates force  $f_m = 8\pi\kappa|m|$
- Example: Spont curvature  $\approx -1/(100 \text{ nm})$  implies  
curvature-induced constriction force  $f_m \approx 25 \text{ pN}$
- Adhesion-induced constriction force  $f_w = 2\pi(2\kappa|W|)^{1/2}$

# ESRCT-induced Fission

- ESCRTs assemble at inner leaflet of cell membrane:

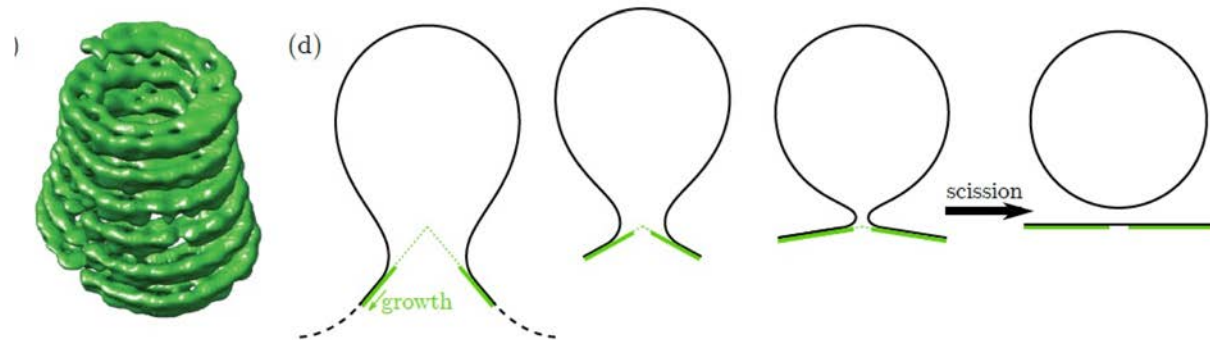
- Domes:

Fabrikant et al,  
PLoS CB (2009)



- Cones:

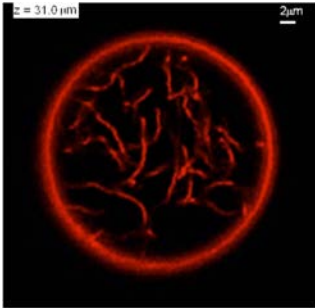
Schöneberg et al,  
Nature RMCB (2017)



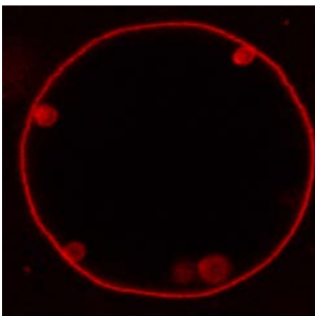
Agudo-Canalejo, RL, (under review)

- Closure via cones is energetically more favorable
- Cones generate constriction force  $f_W \approx 100$  pN

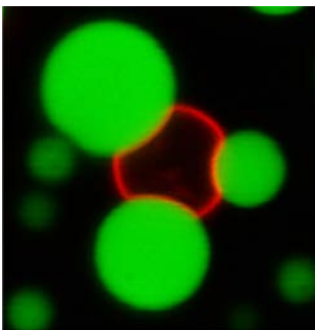
# Forces Generated by Membranes



- Spontaneous Tubulation:  
Spont curvature generates spont tension  
and constriction forces



- ESCRT-induced budding and fission:  
Adhesion-induced constriction forces

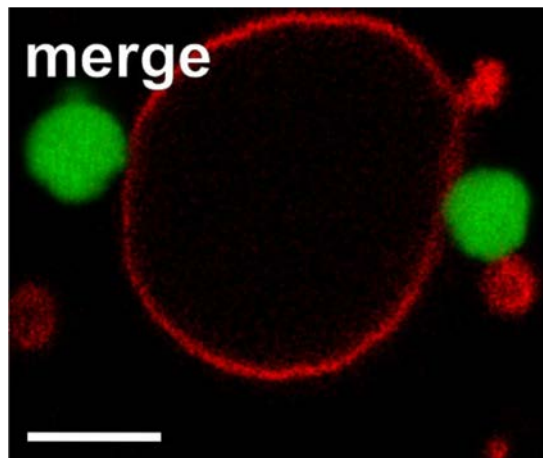


- Interactions with membrane-less organelles:  
Capillary forces and curvature generation

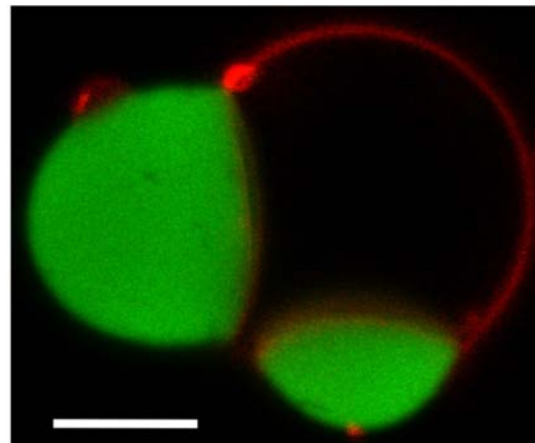
# Membrane-less Organelles

Brangwynne ... Hyman, *Science* (2009)

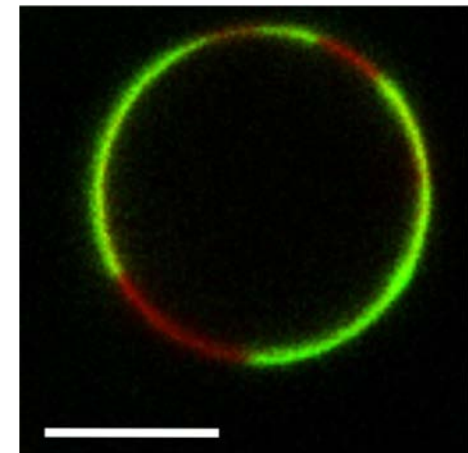
- Membrane-less organelles that behave like liquid droplets
- Enriched in intrinsically disordered proteins (IDPs)
- Example for IDP: RNA-binding protein FUS
- Interaction of FUS-droplets with GUVs, two subsequent wetting transitions:



dewetting for  
high salt



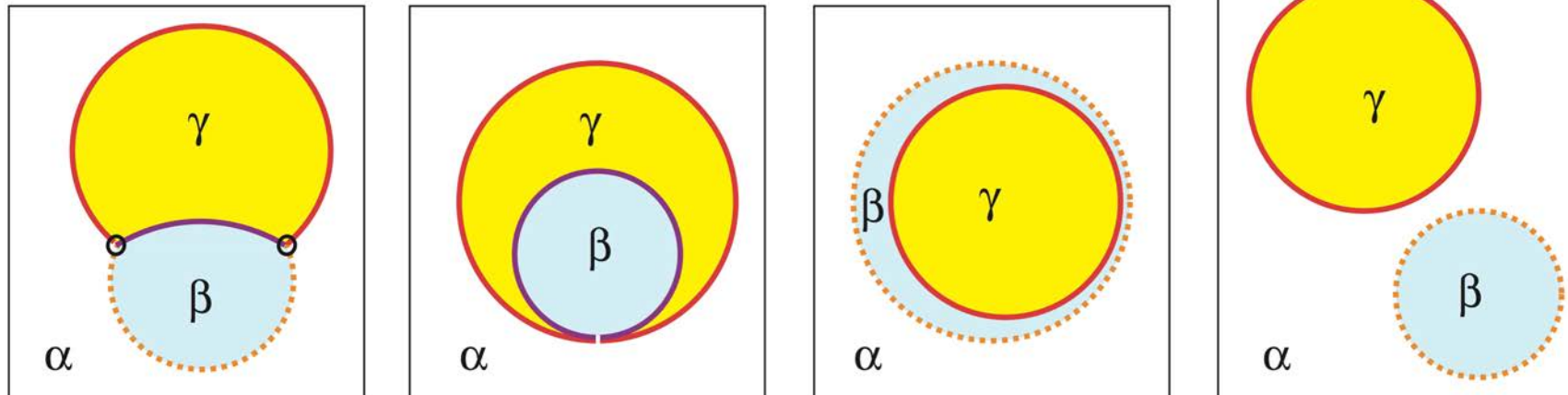
partial wetting for  
intermediate salt



complete wetting  
for low salt

# Shaping by Capillary Forces

RL, J Phys Chem B (Febr, 2018)



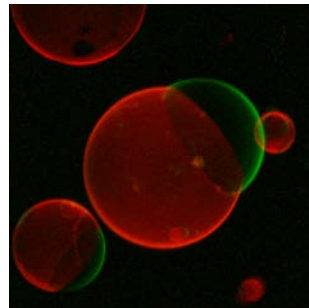
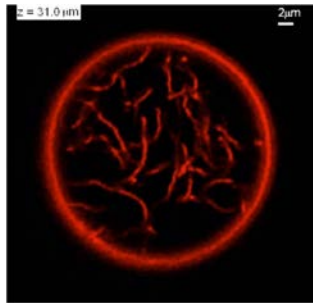
- Apparent and intrinsic contact angles
- Spherical membrane segments imply relation

$$M_{\gamma\alpha} \left( \frac{\Sigma_{\gamma\alpha}^{\text{eff}}}{\Sigma_{\alpha\beta}} - \frac{\sin \theta_{\beta}^{\text{ap}}}{\sin \theta_{\gamma}^{\text{ap}}} \right) = M_{\gamma\beta} \left( \frac{\Sigma_{\gamma\beta}^{\text{eff}}}{\Sigma_{\alpha\beta}} - \frac{\sin \theta_{\alpha}^{\text{ap}}}{\sin \theta_{\gamma}^{\text{ap}}} \right)$$

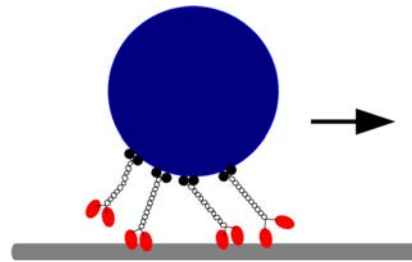
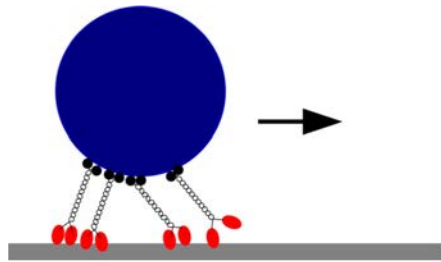
- Effective membrane tensions involve spont curv



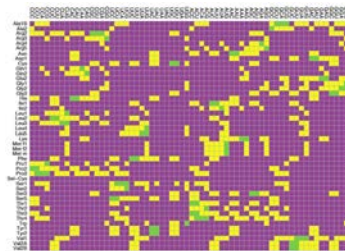
# Basic Modules for Synthetic Cells



- Membrane and vesicles, fluid compartments, remodeling

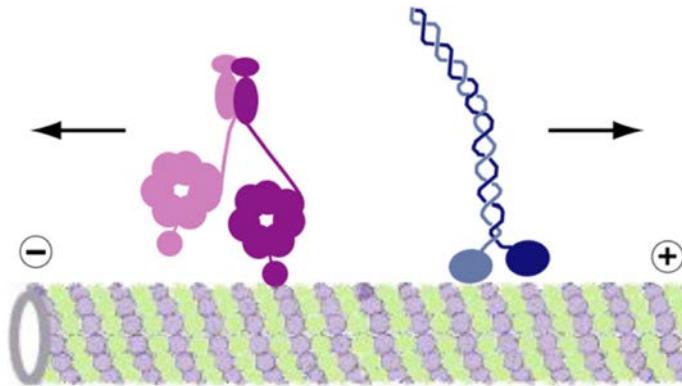


- Directed transport by molecular motors, free energy transduction

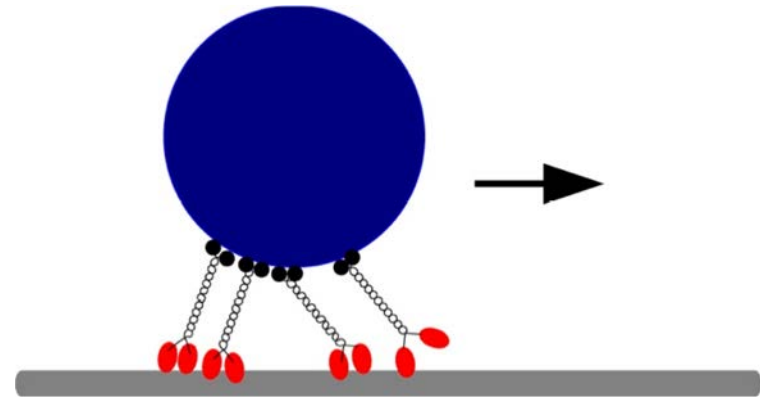


- Template-directed assembly, ribosomes, protein synthesis

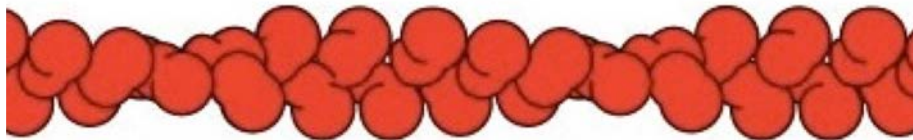
# Biomolecular Machines



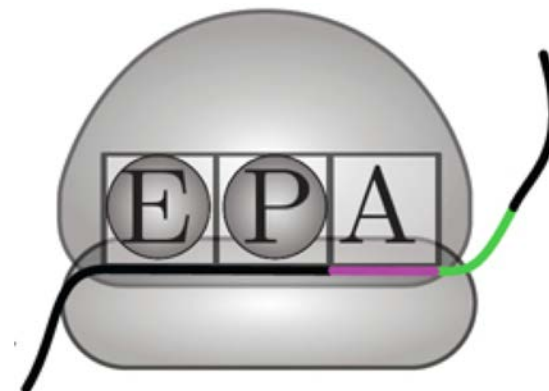
- Intro: Stepping motors



- Transport: Motor teams



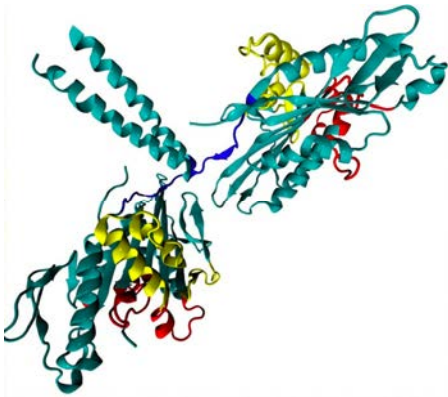
- Structural remodelling:  
Actin filaments



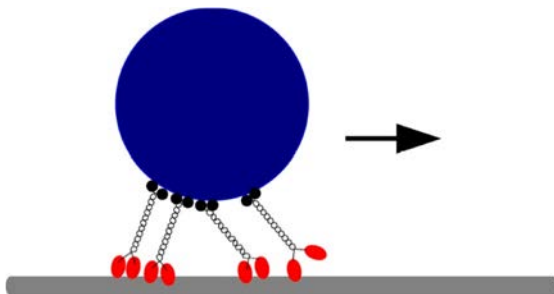
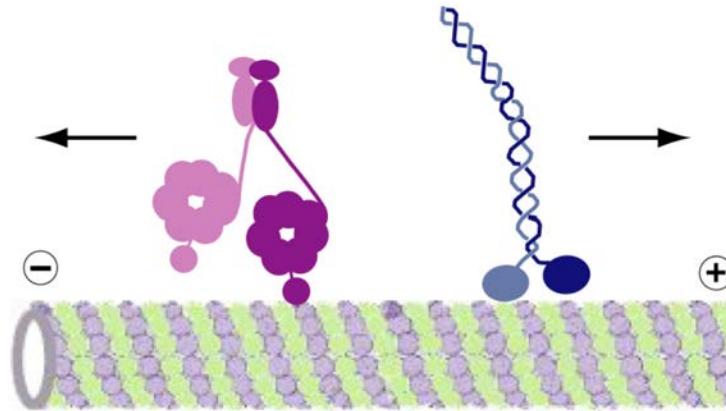
- Information processing:  
Ribosomes

# Multiscale Aspects of Mol Motors

- ATP hydrolysis  $\sim 1$  nm



- Mechanical steps  $\sim 10$  nm



- Cargo transport by motor teams  $\sim 100$   $\mu$ m

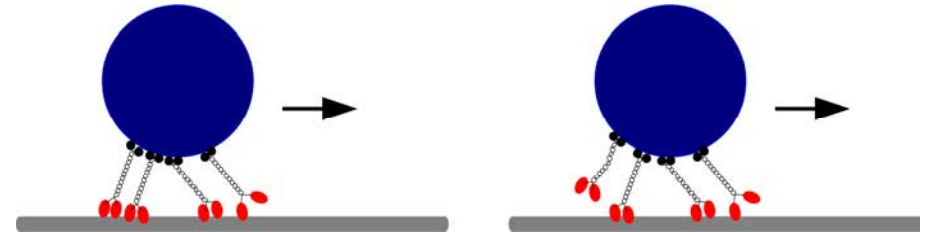


- Traffic of many motors/cargos and phase transitions

# Cargo Transport by Motor Teams

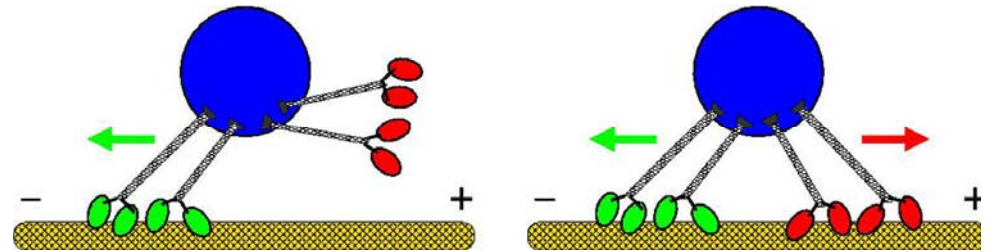
- Transport by  $N$  identical motors

Klumpp and RL, *PNAS* (2005)



- Transport by two antagonistic motor teams,  
Stochastic tug-of-war

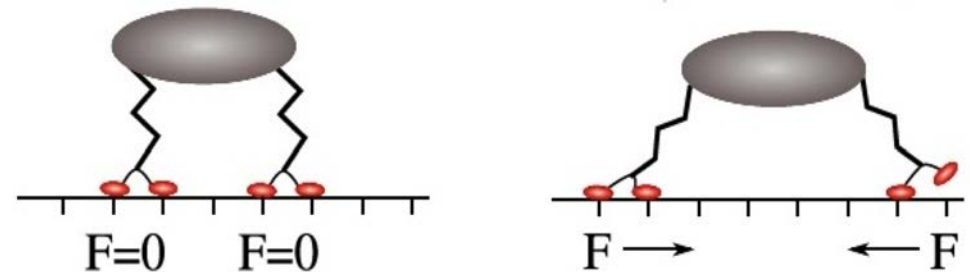
M. Müller et al, *PNAS* (2008)



- Elastic linkers between motors and cargo

Berger et al, *PRL* (2012)

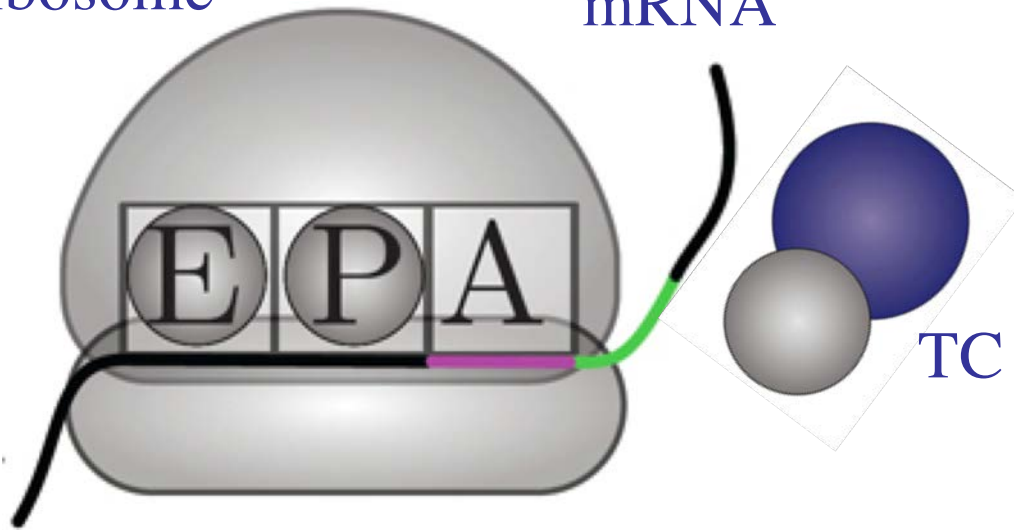
Ucar, RL, *Soft Matter* (2017)



# Protein Synthesis by Ribosomes

Ribosome

mRNA



TC = ternary complex =  
tRNA + EF-Tu + GTP

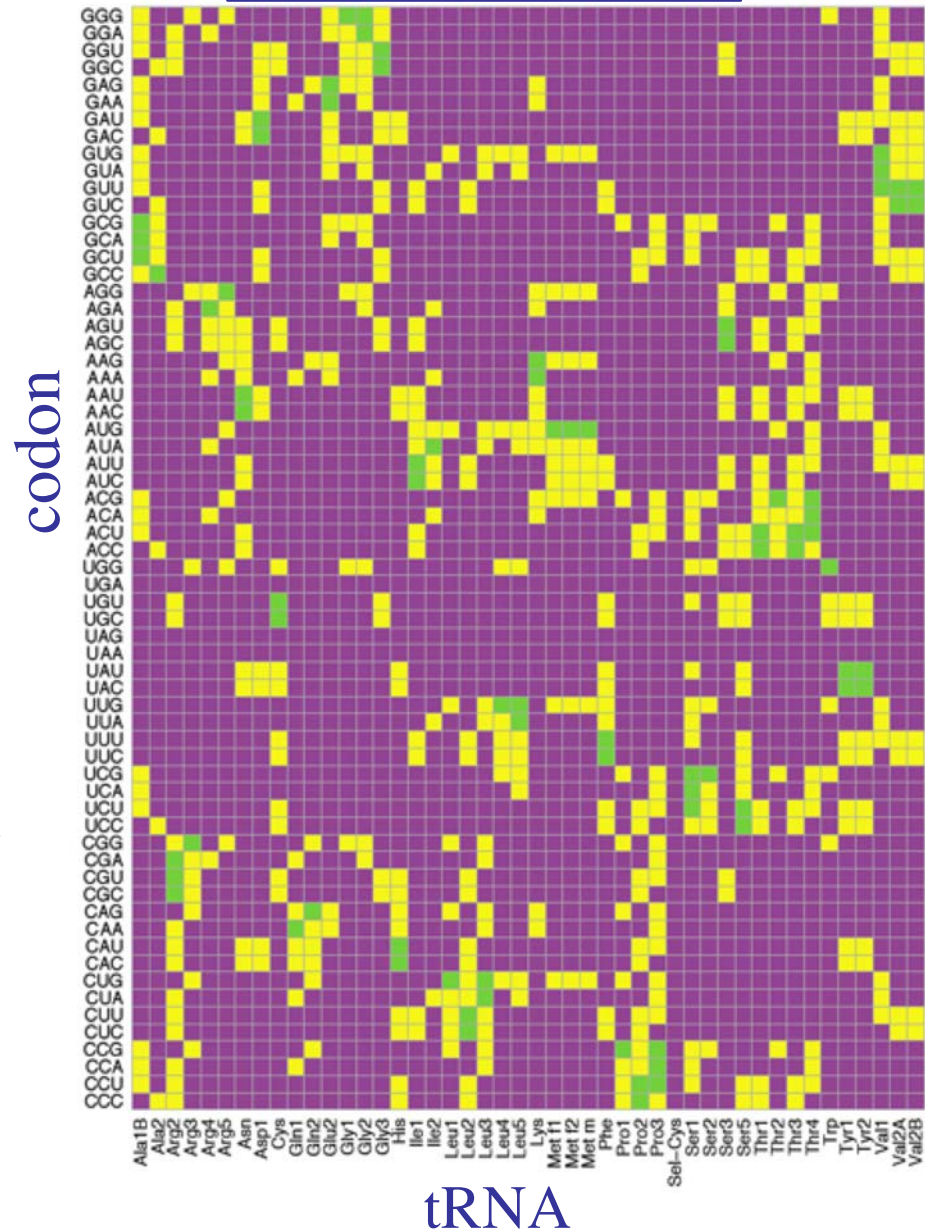
EF-Tu =  
most abundant protein

- Ribosome steps along codons of mRNA (purple -> green) consuming one ternary complex at each codon
- Elongation cycle during one step:
  - Decoding of codon by binding/accommodation of tRNA
  - Elongation of growing peptide chain by one amino acid
  - Translocation of mRNA together with two tRNAs

# Codon-tRNA Relationships

- red/purple = non-cognate  
released after initial binding
- yellow = near-cognate  
decoding => wrong amino acid
- green = cognate  
decoding => correct amino acid
- ‚Ocean‘ of non-cognates  
with some near-cognates  
and a few cognates

## Decoding pattern



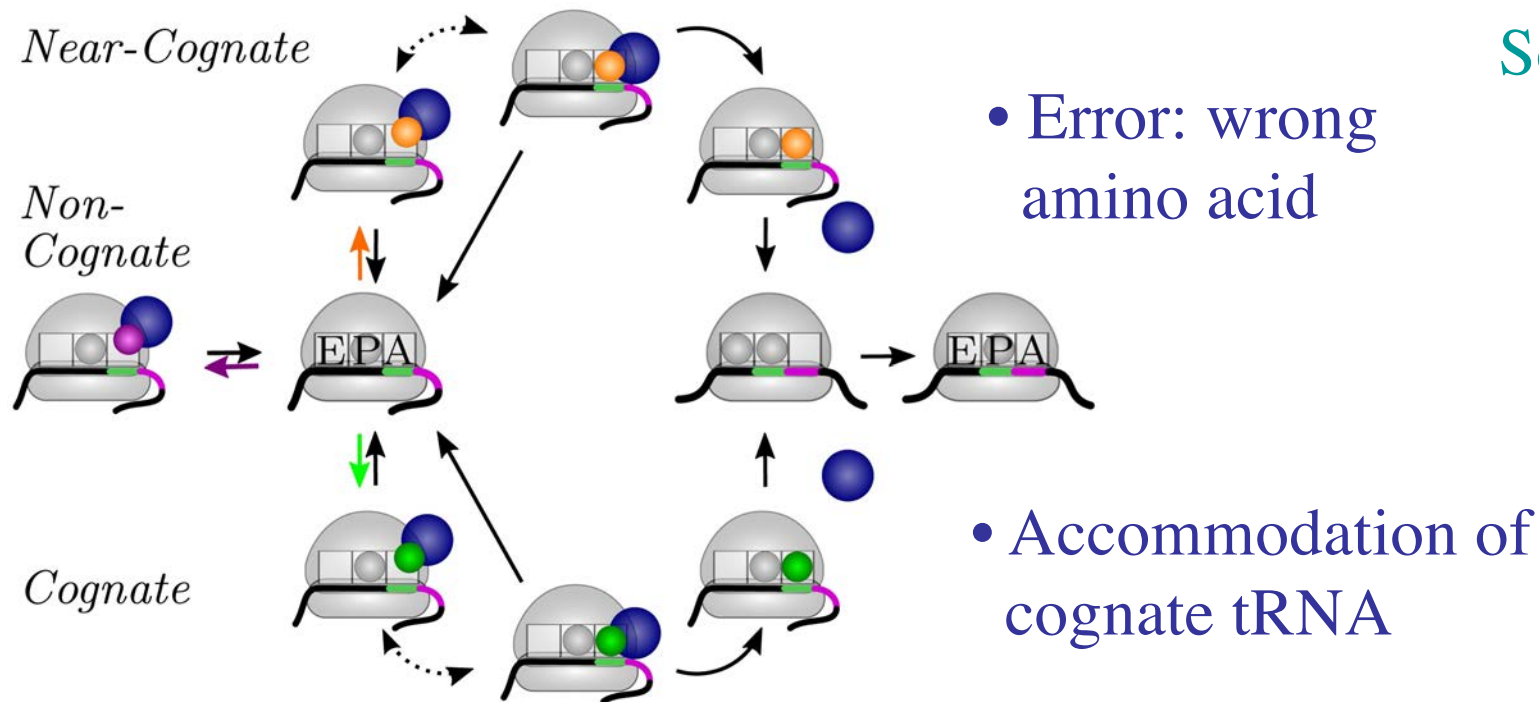
# Single Elongation Cycle

Rudorf, Thommen, Rodnina, RL,  
*PLoS Comp Biol* (2014)

- Three branches for tRNA binding:



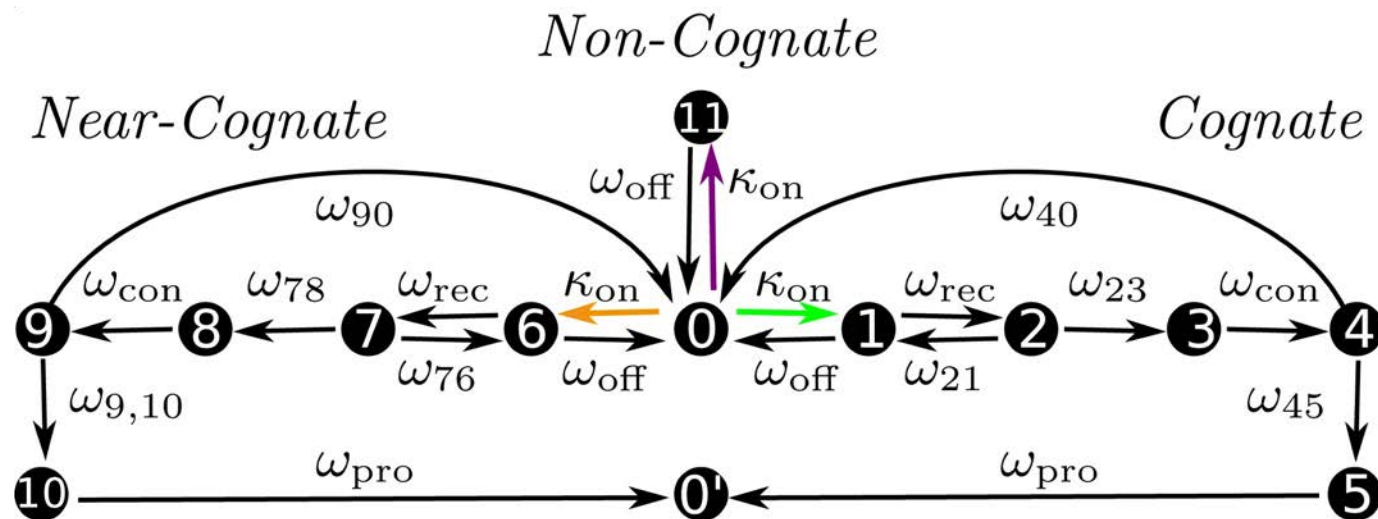
Sophia Rudorf



- **Competition** between cognate, near-cognate, and non-cognate tRNAs

# Markov Process

- Map cartoon of multistep process onto Markov chain:



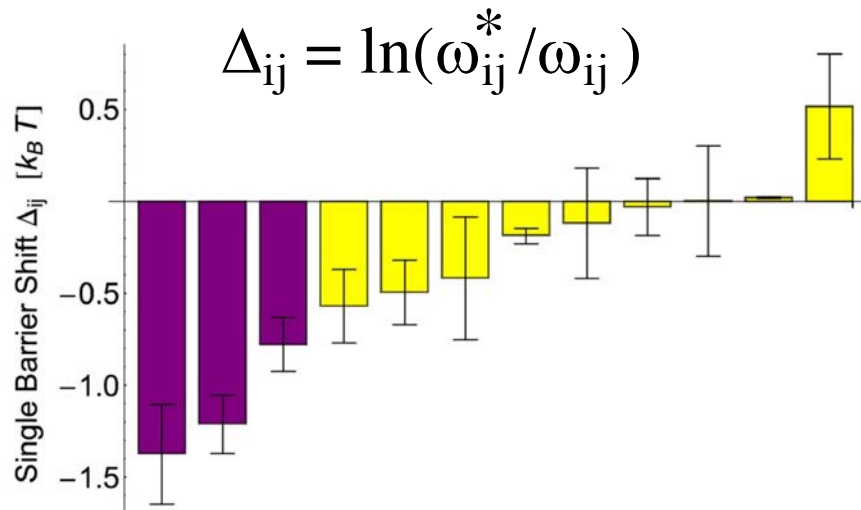
- Individual transitions:  
initial binding, recognition, initial selection, GTP hydrolysis, phosphate release, proof reading, full accommodation
- All transition rates  $\omega_{ij}$  have been measured in vitro
- Some rates identical for both cognates and near-cognates



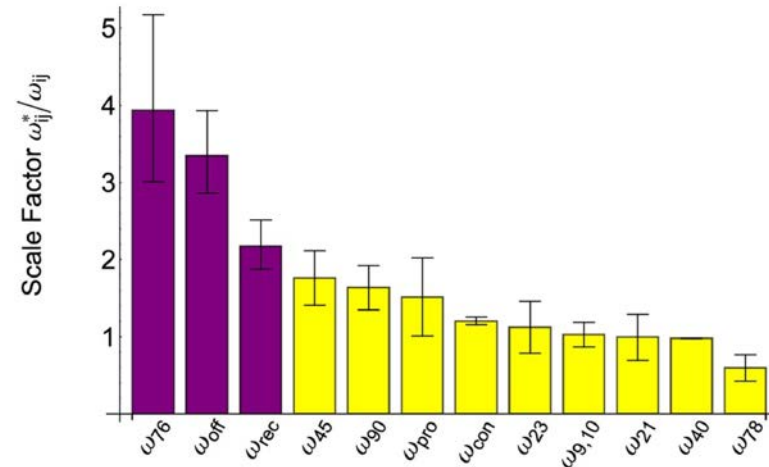
# From In-Vitro to In-Vivo Rates

Rudorf, Thommen, Rodnina, RL, *PLoS Comp Biol* (2014)

- Single barrier shifts



- Scale factors  $\omega_{ij}^*/\omega_{ij}$

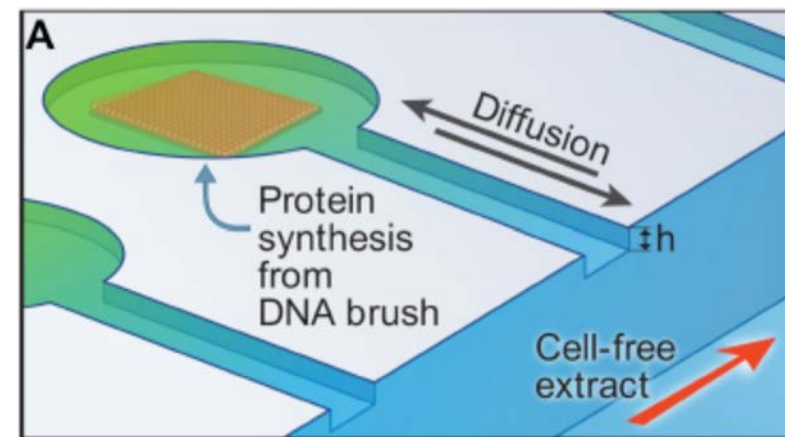
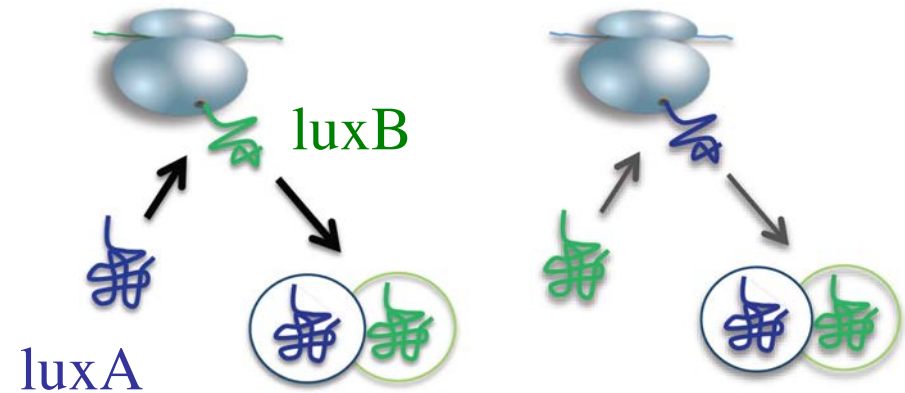


- Three in-vivo rates (purple) are significantly increased:
  - rejection rate  $\omega_{76}$  for near cognates
  - dissociation rate  $\omega_{off}$  after initial binding
  - recognition rate  $\omega_{rec}$  for cognates and near-cognates

# Assembly of Protein Complexes

Shieh ... Kramer, Bukau, *Science* (2015)

- Co-translational assembly:  
*in vivo* synthesis of two proteins,  
assembly during translation,  
luxA binds to emerging luxB
- Protein synthesis on a chip:  
different DNA compartments  
for different proteins,  
control of spatial separation  
between different compartments

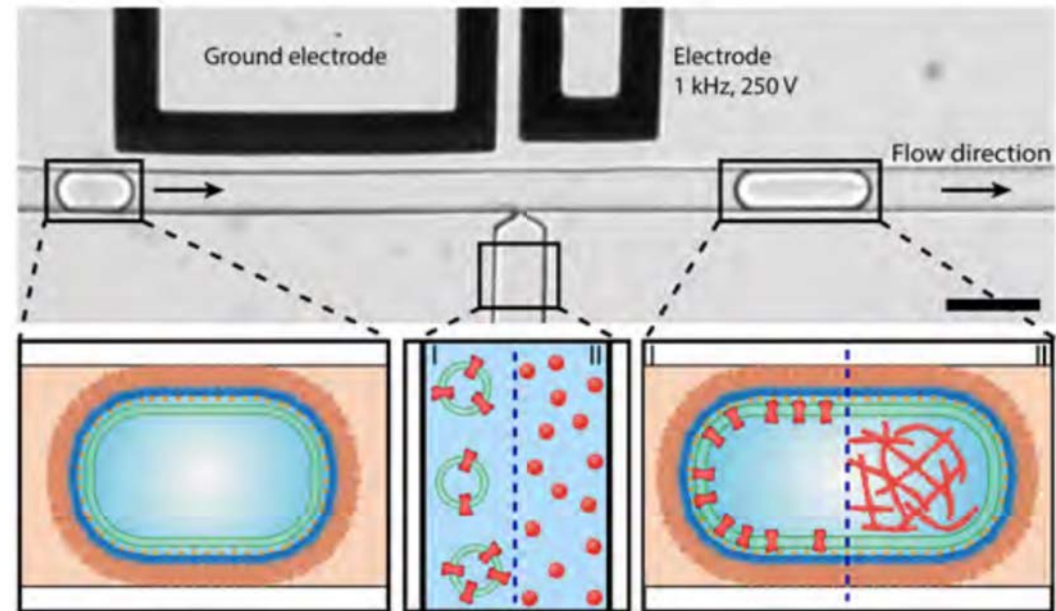
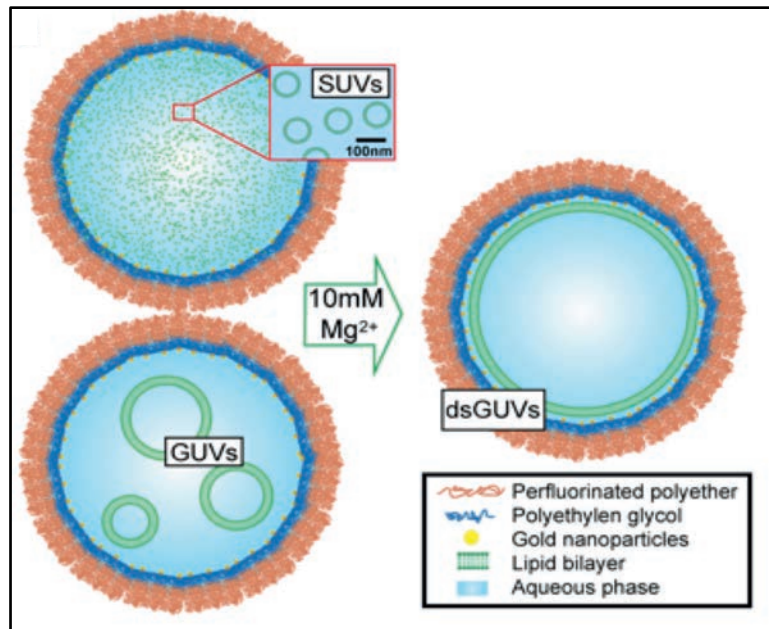


Karzbrun ... Bar-Ziv, *Science* (2014)

# Sequential Bottom-Up Assembly

Weiss et al, *Nature Materials* (Nov. 2017)

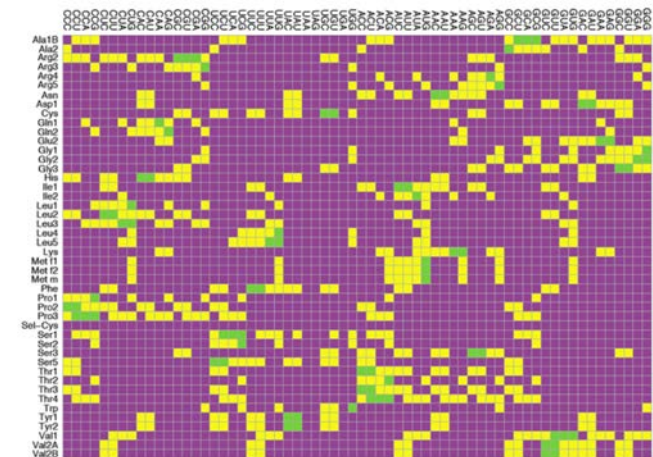
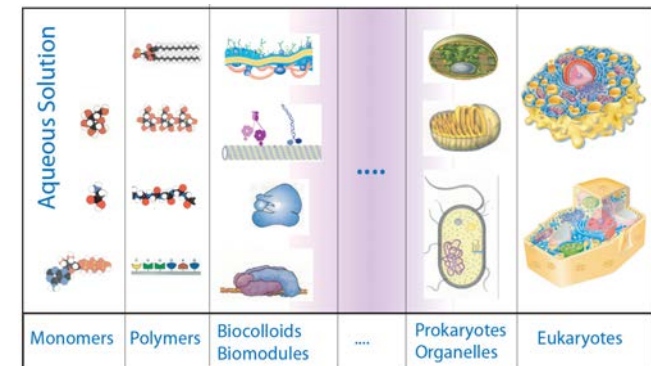
- Four MPIs within MaxSynBio, leading PI: **Joachim Spatz**



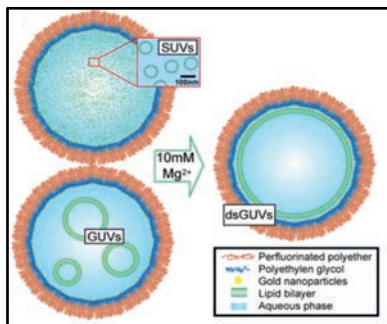
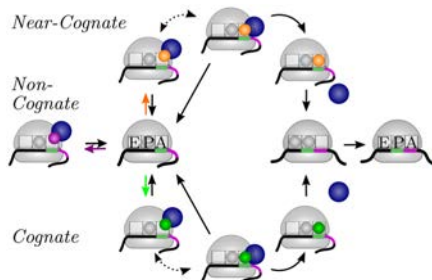
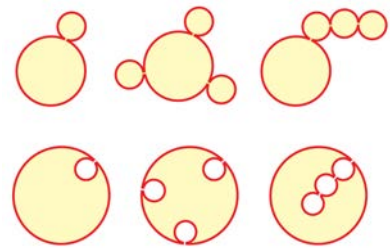
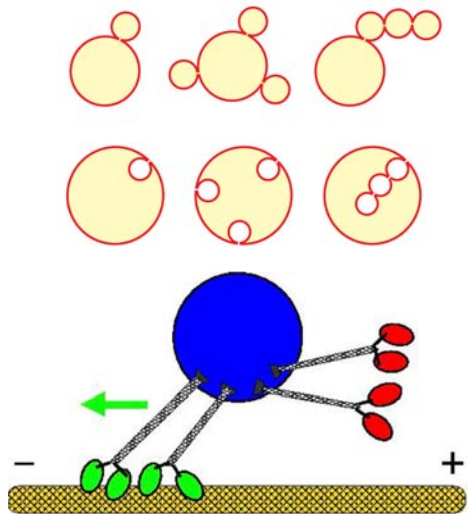
- Water-in-Oil emulsion droplets
- Formation of GUV supported by the droplet surface
- Additional components by pico-injection
- Example: ATP synthase

# Perspectives and Challenges

- Further steps of sequential assembly:  
Compartments + ATP synthase + filaments + motors + ...
- Importance of ionic conditions
- Unwanted interactions, complexes
- Alternative assembly pathways
- Evolution via selection (failures)
- Evolution as a learning process
- Ancestor cells after  $\sim 10^8$  years
- Synthetic cells after ??? years
- Persistent complexity gap ?



# Summary



- Membrane compartments, multiresponsive, many architectures
- Molecular motors, cargo transport and concentr gradients
- Protein synthesis, comparison of in vivo and in vitro
- Droplet-stabilized GUVs, new platform for sequential assembly

# Coworkers



- Membranes

Rumiana Dimova  
Tom Robinson  
Jaime Agudo-Canalejo  
Tripta Bhatia  
Yunuen Avalos Padillo  
Jan Steinkühler

- Motors + Ribosomes

Stefan Klumpp  
Sophia Rudorf  
Mehmet Ucar  
Stefanie Foerste  
Nadin Haase  
Simon Christ

- Collaborations

Marina Rodnina  
Joachim Spatz  
Tony Hyman  
Titus Franzmann  
Günther Kramer  
Roy Bar-Ziv