# Critical Particle Sizes for the Engulfment of Nanoparticles by Membranes and Vesicles with Bilayer Asymmetry – Supporting Information

Jaime Agudo-Canalejo and Reinhard Lipowsky\*

Theory & Biosystems, Max Planck Institute of Colloids and Interfaces, 14424 Potsdam, Germany

E-mail: lipowsky@mpikg.mpg.de

This Supporting Information contains technical details about

- A. Free energy landscapes for enguliment process;
- B. Exocytic engulfment of nanoparticles; and
- C. Spreading dynamics and engulfment rate.

It also contains the three supplementary figures S1 - S3

## A. Free energy landscapes for enguliment process

Geometry of bound and unbound membrane segments. In order to minimize the free energy of the vesicle-particle system, we decomposed the membrane into a bound segment and an unbound segment as shown in Fig. S1 for endocytic engulfment. The bound membrane segment (red in Fig. S1) is in contact with the nanoparticle of radius  $R_{\rm pa}$  and extends up to the contact line which defines the wrapping angle  $\phi$ . This angle varies from

<sup>\*</sup>To whom correspondence should be addressed

 $\phi = 0$  for the free particle state to  $\phi = \pi$  for the completely engulfed state and can be regarded as the reaction coordinate for the engulfment process. The total membrane area  $A = 4\pi R_{\rm ve}^2$  is equal to the sum of the area  $A_{\rm bo}$  of the bound membrane segment and the area  $A_{\rm un}$  of the unbound segment.

The bound segment of the vesicle membrane follows the contour of the particle, and thus assumes the shape of a spherical cap with mean curvature  $M = \pm 1/R_{\text{pa}}$  where the plus and minus sign correspond to the endocytic and exocytic process, respectively. The area of the bound segment is given by

$$A_{\rm bo} = 2\pi R_{\rm pa}^2 \left(1 - \cos\phi\right). \tag{S1}$$

If we cut the spherical particle along the contact line, we obtain two spherical caps. The spherical cap adjacent to the bound membrane segment has the volume

$$V_{\rm bo} = \frac{4\pi}{3} R_{\rm pa}^3 (2 + \cos\phi) \left[\sin(\phi/2)\right]^4 \,. \tag{S2}$$



Figure S1: Vesicle membrane (red-blue) in contact with a spherical nanoparticle (gray) of radius  $R_{\rm pa}$ . The vesicle shape is axially symmetric with respect to the vertical dashed line. The wrapping (or spreading) angle  $\phi$  denotes the position of the contact line which partitions the membrane into a bound (red) and an unbound (blue) segment. The wrapping angle varies from  $\phi = 0$  for the onset of adhesion up to  $\phi = \pi$  for the completely engulfed state. The bound and unbound membrane segment have the areas  $A_{\rm bo}$  and  $A_{\rm un} = A - A_{\rm bo}$ , respectively.

The unbound membrane segment does not experience molecular interactions with the particle and its shape is determined (i) by the location of the contact line, which provides the circular boundary of the unbound membrane segment, (ii) by the area  $A_{\rm un} = A - A_{\rm bo}$  of the unbound segment, (iii) by the effective volume  $V \pm V_{\rm bo}$ , which is enclosed by the unbound membrane segment and the additional planar surface that spans the circular contact line, where the plus and minus sign applies to endo- and exocytosis, respectively; and (iv) by the spontaneous curvature m.

**Decomposition of total free energy.** As explained in the *Methods* section, the total free energy E is equal to the sum of the membrane's bending free energy  $E_{\rm be}$  (main text, Eq. 22) and the adhesion free energy  $E_{\rm ad} = -|W| A_{\rm bo}$ . The total free energy can also be decomposed according to

$$E = E_{\rm bo} + E_{\rm un} \tag{S3}$$

with the free energy

$$E_{\rm bo} \equiv \left[-2\pi |W| R_{\rm pa}^2 + 4\pi\kappa (1\pm mR_{\rm pa})^2\right] \left[1 - \cos(\phi)\right].$$
(S4)

of the bound membrane segment, where the plus and minus sign correspond to endocytic and exocytic engulfment, respectively, and the free energy

$$E_{\rm un} = \int dA_{\rm un} \, 2\kappa (M-m)^2 \tag{S5}$$

of the unbound membrane segment where the integral extends over the area  $A_{\rm un} = A - A_{\rm bo}$ of the unbound segment.

In Eq. S4, we used the convention that the upper and lower signs of the  $\pm$  symbol correspond to the endocytic and exocytic case, respectively. The same convention will be used below in all equations in which a  $\pm$  symbol appears.

Free energy minimization. In order to find the shape of the unbound segment that minimizes  $E_{un}$  for a given value of the wrapping angle  $\phi$  and, thus, for a given location of the contact line, we minimize the shape functional

$$F_{\rm un} \equiv E_{\rm un} + \Sigma (A - A_{\rm bo}) - \Delta P (V \pm V_{\rm bo}) \tag{S6}$$

where  $\Sigma$  and  $\Delta P$  are Lagrange multipliers which ensure that the membrane area has the prescribed value A and that the unbound and bound membrane together enclose the vesicle volume V. The auxiliary volume  $V \pm V_{\text{bo}}$  is bounded by the unbound membrane segment and the additional planar surface that spans the circular contact line. When we calculate the free energy  $E_{\text{un}}(\phi)$  of the unbound membrane segment by minimizing the shape functional  $F_{\text{un}}$  in Eq. S6 for many different values of  $\phi$  within the interval  $0 < \phi < \pi$ , keeping both the area A and the volume V fixed, we obtain the corresponding free energy landscape

$$E(\phi) = \left[-2\pi |W| R_{\rm pa}^2 + 4\pi\kappa (1 \pm mR_{\rm pa})^2\right] \left[1 - \cos(\phi)\right] + E_{\rm un}(\phi)$$
(S7)

where the first term on the right hand side follows from Eq. S4. Typical free energy landscapes  $E(\phi)$  obtained in this way are displayed in Fig. S2 corresponding to the parameter values marked with green diamonds in Fig. 3c. In general, the free energy landscapes may contain additional minima corresponding to additional intermediate states. For the uniform membranes considered in the main text, these additional states can always be ignored because they represent satellite minima very close to the free or completely engulfed states, from which they are separated by tiny energy barriers that can be easily overcome by thermal fluctuations.



Figure S2: Free energy landscapes  $\Delta E(\phi) \equiv E(\phi) - E(0)$  corresponding to the six parameter values marked by green diamonds in Fig. 3c: (a) For the free (or non-engulfment) regime  $\mathcal{F}_{st}$ , the landscape has a minimum at  $\phi = 0$ , which corresponds to the free state  $\mathcal{F}$ , and a maximum at  $\phi = \pi$ , which defines the completely engulfed state  $\mathcal{C}$ ; (b) For the partial engulfment regime  $\mathcal{P}_{st}$ , the landscape has maxima both at  $\phi = 0$  and at  $\phi = \pi$  and a minimum at an intermediate  $\phi$ -value corresponding to a partially engulfed state; (c) For the complete engulfment regime  $\mathcal{C}_{st}$ , the landscape exhibits a minimum at  $\phi = \pi$  and a maximum at  $\phi = 0$ ; (d, e, f) Three landscapes within the bistable regime  $\mathcal{B}_{st}$  with two local minima at  $\phi = 0$  and  $\phi = \pi$  separated by a free energy barrier. In panels (d) and (f), the global minima are provided by the states  $\mathcal{F}$  and  $\mathcal{C}$ , respectively. Panel (e) corresponds to the transition line  $L_*$  at which both states  $\mathcal{F}$  and  $\mathcal{C}$  have the same free energy.

Free energy landscape close to free particle state  $\mathcal{F}$ . Close to the free particle state  $\mathcal{F}$  with  $\phi = 0$ , the free energy landscape behaves as

$$E(\phi) \approx E(0) + \frac{1}{2} E'' \phi^2 \quad \text{with} \quad E'' \equiv \left. \frac{d^2 E(\phi)}{d \phi^2} \right|_{\phi=0}.$$
 (S8)

The free particle state is (meta)stable as long as E'' > 0 and unstable for E'' < 0, which implies that E'' = 0 determines the instability line  $L_{\rm fr}$ . Using the decomposition  $E = E_{\rm bo} + E_{\rm un}$  of the free energy into the contributions from the bound and unbound membrane segments, as in Eqs. S3 - S5, we obtain  $E'' = E''_{\rm bo} + E''_{\rm un}$  with

$$E_{\rm bo}'' = -2\pi R_{\rm pa}^2 |W| + 4\pi\kappa (1 \pm R_{\rm pa} m)^2 \tag{S9}$$

with the plus (minus) sign corresponding to endocytosis (exocytosis).

The instability line  $L_{\rm fr}$  is now determined by

$$E'' = -2\pi R_{\rm pa}^2 |W| + 4\pi \kappa (1 \pm R_{\rm pa} m)^2 + E''_{\rm un} = 0.$$
 (S10)

Alternatively, we may also determine  $L_{\rm fr}$  from the instability relation

$$M_{\rm co} = M_{\rm ms}$$
 or  $\sqrt{R_{\rm pa}^2 |W|/(2\kappa)} = 1 \pm R_{\rm pa} M_{\rm ms}$  (S11)

where the expression for the contact mean curvature  $M_{co}$  as in Eq. 23 of the main text has been used. The two relationships as given by Eqs. S10 and S11 are only equivalent if the unbound membrane segment makes the contribution

$$E''_{\rm un} = 4\pi\kappa R_{\rm pa}[M_{\rm ms} - m] \left[\pm 2 + R_{\rm pa}(M_{\rm ms} + m)\right]$$
(S12)

to the second derivative E''. A combination of  $E''_{bo}$  in Eq. S9 and  $E''_{un}$  in Eq. S12 then leads to

$$E'' = E''_{\rm bo} + E''_{\rm un} = -2\pi R_{\rm pa}^2 |W| + 4\pi\kappa (1 \pm R_{\rm pa} M_{\rm ms})^2 \,. \tag{S13}$$

For endocytosis (+ sign), this relationship is identical to Eq. 6 in the main text.

Relation between membrane area and mechanical membrane tension. In the theoretical approach used here, the mechanical tension  $\Sigma$  does not represent an independent parameter but plays the role of a Lagrange multiplier  $\Sigma$ , see Eq. S6, which is determined in terms of the other parameters in order to ensure that the membrane area has the prescribed value A.<sup>1-3</sup> The minimization procedure typically leads to Lagrange multipliers  $\Sigma$  that correspond to relatively small tensions of the order of  $\kappa/R_{ve}^2$ ,  $\kappa m/R_{ve}$ , or  $\kappa m^2$ . If we stretched the membrane with such a tension, the change in membrane area arising from the membrane's area compressibility would be rather small which provides a consistency check on the theory. In fact, even in the presence of relatively large tensions of the order of 1 mN/m, the membrane area can only change by a few percent without rupturing. Thus, as long as the membrane does not rupture, its area remains constant to a very good approximation. For giant unilamellar vesicles (GUVs), the membrane area A can be directly measured and it is then possible to corroborate the theory by a systematic comparison of calculated and experimentally observed membrane shapes as has been successfully done for lipid vesicles in the absence of nanoparticles.<sup>4-6</sup>

An alternative theoretical approach has been used in Refs. 7 and 8 where the engulfment of nanoparticles was theoretically studied in the presence of a certain prescribed membrane tension  $\Sigma'$ . This tension was treated as an independent control parameter and then represents a 'chemical potential' for membrane area which would govern the exhange of area with a putative area reservoir, in analogy to a grand-canonical ensemble. This approach is motivated by the view that eukaryotic cells control the tension of their plasma membranes, presumably by regulating the osmotic conditions and by remodelling the cytoskeletal forces acting on the cell membrane, which leads to the so-called cortical tension. Because the mechanisms underlying this tension are complex and poorly understood, it is appealing to reduce this complexity to a single tension parameter. One difficulty with this approach is that the measured tension values are quite variable and change during the cell cycle. Indeed, recent experiments provide  $\Sigma'$ -values in the range between 0.05 and 2 mN/m.<sup>9,10</sup> Another difficulty is that the actin-myosin cortex exerts complex patterns of forces onto the cell membrane which contribute to the membrane tension but, at the same time, directly affect the membrane shape, and it is not obvious to what extent these two effects of the cortical forces may be disentangled.

The lipid membranes and vesicles considered in our study do not involve an area (or lipid) reservoir which implies that the membrane area A rather than the membrane tension should be regarded as the basic control parameter. Furthermore, the instability relations for the free and completely engulfed particle states as derived here depend only on *local* 

properties of the membrane close to the nanoparticle and can, thus, also be applied to cell membranes. In fact, one can show that the free energy landscapes for particle engulfment remain unaffected by small tensions  $\Sigma' \ll \Sigma_o \equiv \kappa/R_{\rm pa}^2$ . For a membrane with a clathrin coat, the bending rigidity is  $\kappa = 10^{-18}$  J as measured in Ref. 11 which leads to crossover tensions  $\Sigma_o \geq 0.4$  mN/m for particle sizes  $R_{\rm pa} \leq 50$  nm as studied experimentally in Refs. 12 and 13.

### **B.** Exocytic engulfment of nanoparticles

Instability relations for exocytic engulfment. For exocytic engulfment, the curvature  $M_{\rm ms}$  of the membrane segment adjacent to the free particle state  $\mathcal{F}$  (main text, Fig. 5) must be smaller than  $1/R_{\rm pa}$  in order to ensure that the membrane and the particle do not intersect each other. Furthermore, the contact mean curvature is given by  $M_{\rm co} = -\frac{1}{R_W} + \frac{1}{R_{\rm pa}}$  as explained in the *Methods* section. As a consequence, the relation  $M_{\rm co} = M_{\rm ms}$  for the instability line  $L_{\rm fr}$  of the free state  $\mathcal{F}$  leads to

$$R_{\rm pa} = R_{\rm fr} \equiv \frac{1}{M_{\rm ms} + R_W^{-1}}$$
 and  $M_{\rm ms} > -1/R_W$  (*L*<sub>fr</sub>, exocytosis). (S14)

and the membrane segment starts to spread over the particle if

$$R_{\rm pa} > R_{\rm fr}$$
 and  $M_{\rm ms} > -1/R_W$  (unstable  $\mathcal{F}$ , exocytosis). (S15)

For strongly curved membrane segments with a negative mean curvature  $M_{\rm ms}$  smaller than  $-1/R_W$ , the free state  $\mathcal{F}$  is stable for all particle sizes, *i.e.*, the critical particle size  $R_{\rm fr} = \infty$ .

For exocytic engulfment, the curvature  $M'_{\rm ms}$  of the mother membrane adjacent to the membrane neck of the completely engulfed state C (main text, Fig. 6) must be larger than  $-1/R_{\rm pa}$  in order to ensure that the mother membrane does not intersect the membrane segment bound to the particle. The instability line  $L_{\rm ce}$  for the state C as determined by  $M_{\rm co} + M'_{\rm ms} = 2m$  now has the form

$$R_{\rm pa} = R_{\rm ce} \equiv \frac{1}{2m + R_W^{-1} - M_{\rm ms}'} \quad \text{for} \quad M_{\rm ms}' < 2m + 1/R_W \quad (L_{\rm ce}, \,\text{exocytosis})\,, \qquad (S16)$$

and the membrane neck starts to open if

$$R_{\rm pa} < R_{\rm ce}$$
 and  $M'_{\rm ms} < 2m + 1/R_W$  (unstable  $\mathcal{C}$ , exocytosis). (S17)

If the unbound membrane segment has a mean curvature  $M'_{\rm ms}$  larger than  $2m + \frac{1}{R_W}$ , the completely engulfed state C is unstable for all particle sizes, *i.e.*,  $R_{\rm ce} = \infty$ . The physical requirement that the membrane has no self-intersections in state C leads to the additional condition that  $M'_{\rm ms}$  is larger than  $-1/R_{\rm pa}$ . Therefore, stable states C without self-intersections are only possible for  $-\frac{1}{R_{\rm pa}} < M'_{\rm ms} < 2m + \frac{1}{R_W}$ .

These instability lines and instability criteria for exocytic engulfment can be transformed into those for endocytic engulfment, if we change (i) the sign of the spontaneous curvature mas well as (ii) the signs of the curvatures  $M_{\rm ms}$  and  $M'_{\rm ms}$  of the two membrane segments. This 'mirror symmetry' implies that we have a one-to-one correspondence between the engulfment diagrams for exocytic and endocytic engulfment as illustrated further below for the case of weakly curved membranes.

For notational simplicity, we have used the same notation  $R_W$  for the adhesion length of both the endocytic and the exocytic process. Note, however, that the two adhesion lengths may have different numerical values because the molecular interactions described by the adhesive strength W may be different on the two sides of the asymmetric bilayer.

Critical particle sizes for exocytic engulfment. The equations of the previous subsection imply that the intersection point of the two instability lines is again located at  $m = \frac{1}{2} (M_{\rm ms} + M'_{\rm ms})$  but that the relative positions of the intermediate size regimes  $\mathcal{B}_{\rm st}$  and  $\mathcal{P}_{\rm st}$  are now swapped compared to the endocytic case. Therefore, the engulfment process



Figure S3: Exocytic engulfment of nanoparticles by weakly curved membranes: Different engulfment regimes  $\mathcal{F}_{st}$ ,  $\mathcal{B}_{st}$ ,  $\mathcal{C}_{st}$  and  $\mathcal{P}_{st}$  as a function of particle size  $R_{pa}$  and spontaneous curvature m, both measured in units of the adhesion length  $R_W$ . (a) Concave membrane segments with small negative curvatures  $M_{ms} = M'_{ms} = -0.05/R_W$ ; (b) Flat membrane segments with vanishing curvatures  $M_{ms} = M'_{ms} = 0$ ; and (c) Convex membrane segments with small positive curvatures  $M_{ms} = M'_{ms} = +0.05/R_W$ . The two instability lines  $L_{fr}$  and  $L_{ce}$  for the free and completely engulfed states are given by Eq. S14 and Eq. S16 and define the critical particle sizes  $R_{fr}$  and  $R_{ce}$ . The bistable regimes  $\mathcal{B}_{st}$  contain the transition lines  $L_*$  (dashed) at which the free and completely engulfed states coexist. All four engulfment regimes meet at the 'multicritical' intersection points of the two instability lines. Compared to endocytic engulfment (main text, Fig. 7), the relative locations of the regimes  $\mathcal{B}_{st}$  and  $\mathcal{P}_{st}$ have been swapped.

is continuous for  $m < \frac{1}{2}(M_{\rm ms} + M'_{\rm ms})$  and discontinuous for  $m > \frac{1}{2}(M_{\rm ms} + M'_{\rm ms})$ , see Table S1. The latter table also contains the two critical particle radii for exocytic engulfment as obtained from the corresponding instability criteria in Eqs. S15 and S17.

Table S1: Critical particle sizes for *exocytic* enguliment as derived from Eqs. S15 and S17.

Range of spontaneous curvature $m$	Intermediate size regime	Engulfment process	Lower critical size	Upper critical size
$m > \frac{1}{2} \left( M_{\rm ms} + M_{\rm ms}' \right)$	bistable $\mathcal{B}_{st}$	discontinuous	$R_{\rm ce}$	$R_{ m fr}$
$m < \frac{1}{2} \left( M_{\rm ms} + M_{\rm ms}' \right)$	partial $\mathcal{P}_{st}$	continuous	$R_{\mathrm{fr}}$	$R_{\rm ce}$

Exocytic engulfment by weakly curved mother membranes. For sufficiently large values of the vesicle size  $R_{\rm ve}$ , the two membrane curvatures  $M_{\rm ms}$  and  $M'_{\rm ms}$  can again be

neglected. The corresponding engulfment diagram is depicted in Fig. S3b as a function of particle size  $R_{\rm pa}$  and spontaneous curvature m, both measured in units of the adhesion length  $R_W$ . Inspection of Fig. S3b shows that exocytic engulfment by flat membranes leads to partially engulfed states for negative spontaneous curvature and to bistability for positive spontaneous curvature. This behavior for the exocytic process is exactly the opposite of the behavior for the endocytic process for which partially engulfed states occur for positive spontaneous curvature and bistability is found for negative spontaneous curvatures (main text, Fig. 7b). For small but finite values of the segment curvatures  $M_{\rm ms}$  and  $M'_{\rm ms}$ , the regimes for exocytic engulfment again undergo small changes, primarily determined by the sign of  $M_{\rm ms} + M'_{\rm ms}$ , as illustrated in Fig. S3a and Fig. S3c.

A detailed comparison of the different regimes for exocytic and endocytic engulfment as displayed in Fig. S3 and Fig. 7 shows that the exocytic diagrams can be obtained from the endocytic ones if we simultaneously change the sign of the spontaneous curvature m as well as the signs of the segment curvatures  $M_{\rm ms}$  and  $M'_{\rm ms}$ . In this way, we obtain Fig. S3c from Fig. 7a and Fig. S3a from Fig. 7c. This 'mirror symmetry' of the engulfment diagrams is a direct consequence of the corresponding 'mirror symmetry' of the instability criteria as pointed out after Eq. S17.

Adhesion length and spontaneous curvature from critical particle sizes. If the exocytic engulfment process is continuous and proceeds *via* partially engulfed states, the two critical particle sizes  $R_{\rm fr}$  and  $R_{\rm ce} > R_{\rm fr}$  are accessible to direct observation, either in experimental or in simulations studies. From the observed critical sizes, we can then determine the adhesion length *via* 

$$R_W = \frac{R_{\rm fr}}{1 - R_{\rm fr} M_{\rm ms}} \quad \text{(cont exocytosis)} \tag{S18}$$

and the spontaneous curvature via

$$m = \frac{1}{2} \left[ \frac{1}{R_{\rm ce}} - \frac{1}{R_{\rm fr}} + M_{\rm ms} + M_{\rm ms}' \right] \quad \text{(cont exocytosis)} \tag{S19}$$

as follows from Eqs. S14 and S16.

#### C. Kinetics of membrane spreading and engulfment rate

Force balance at the contact line. The spreading of the membrane over the particle surface is induced by the attractive membrane-particle forces and proceeds *via* the displacement of the contact line. For the engulfment of a spherical particle, the membrane geometry is axially symmetric and the contact line has the total length

$$L_{\rm co} = 2\pi R_{\rm pa} \,\sin(\phi)\,. \tag{S20}$$

The position of the contact line is determined by the contact point of the membrane contour. The coordinate of this point is taken to be the arc length

$$s \equiv R_{\rm pa}\,\phi\tag{S21}$$

of the bound membrane contour measured from the south pole of the particle with  $\phi = 0$  as in Fig. S1. The displacement of the contact line now corresponds to changes in s.

The contact line at position s experiences two forces: a thermodynamic driving force and a friction force. The thermodynamic driving force  $F_1$  reflects the change in the system's energy as we displace the contact line. This force has the form

$$F_1 = -\frac{d E_s(s)}{d s} = -\frac{1}{R_{\rm pa}} \frac{d E(\phi)}{d \phi}$$
(S22)

where  $E_s(s)$  is the free energy landscape of the system as a function of s, *i.e.*,  $E_s(s) =$ 

 $E(\phi(s)) = E(s/R_{\rm pa})$  with the free energy landscape  $E(\phi)$  as discussed before, see Fig. S2. The force  $F_2$ , on the other hand, depends on the dissipation mechanism. If the contact line does not move, there will be no friction. Therefore, the friction force  $F_2$  is taken to be proportional to the velocity v = ds/dt of the contact line which is equal to the derivative of arc length s with respect to time t. The friction coefficient for the displacement of the whole contact line should be proportional to the length  $L_{\rm co}$  of the contact line which implies  $F_2 = \eta_{\rm eff} L_{\rm co} ds/dt$  which defines the effective dynamic viscosity  $\eta_{\rm eff}$ . Using Eq. S20 for the contact length  $L_{\rm co}$  and changing variables from arc length s to wrapping angle  $\phi$  via  $s = R_{\rm pa} \phi$ , we obtain the  $\phi$ -dependent friction force

$$F_2 = 2\pi \eta_{\text{eff}} R_{\text{pa}}^2 \sin(\phi) \frac{d\phi}{dt} \,. \tag{S23}$$

We now balance the thermodynamic driving force  $F_1$  in Eq. S22 with the friction force  $F_2$  in Eq. S23, *i.e.*, we set  $F_1 = F_2$  which leads to the equation of motion for the contact line as given by

$$\sin(\phi) \frac{d\phi}{dt} = -\frac{1}{2\pi \eta_{\text{eff}} R_{\text{pa}}^3} \frac{dE(\phi)}{d\phi}$$
(S24)

which is identical with Eq. 17 in the main text.

Size-dependent engulfment rate. The equation of motion for the contact line (Eq. S24) involves the gradient  $dE/d\phi$  of the free energy landscape which can again be decomposed into two contributions from the bound and unbound membrane segment, *i.e.*,

$$\frac{d E(\phi)}{d \phi} = \frac{d E_{\rm bo}(\phi)}{d \phi} + \frac{d E_{\rm un}(\phi)}{d \phi}$$
(S25)

with

$$\frac{d E_{\rm bo}(\phi)}{d \phi} = \left[-2\pi |W| R_{\rm pa}^2 + 4\pi \kappa_{\rm bo} (1 \pm m_{\rm bo} R_{\rm pa})^2\right] \sin(\phi)$$
(S26)

with the plus sign corresponding to endocytosis.

As far as the gradient  $dE_{\rm un}(\phi)/d\phi$  of the unbound membrane (or mother membrane) is concerned, it is intuitively plausible that  $E_{\rm un}(\phi)$  changes primarily by shape changes of the unbound membrane segment close to the contact line and that these changes are small if this segment can adapt its mean curvature to the spontaneous curvature m. For m = 0, for example, this segment can attain a shape close to a catenoid which has vanishing bending energy and, thus, makes no contribution to  $E_{\rm un}$ . This expectation can be directly confirmed for the initial spreading close to the free state  $\mathcal{F}$ , *i.e.*, for small values of the wrapping angle  $\phi$  because

$$\frac{d E_{\rm un}(\phi)}{d \phi} \approx E_{\rm un, fr}'' \phi = 4\pi \kappa R_{\rm pa}[M_{\rm ms} - m] \left[\pm 2 + R_{\rm pa}(M_{\rm ms} + m)\right] \phi \quad \text{for small } \phi \qquad (S27)$$

as follows from  $E_{\rm un}(\phi) \approx E_{\rm un}(0) + \frac{1}{2} E_{\rm un,fr}'' \phi^2$  and Eq. S12 with  $E_{\rm un}'' = E_{\rm un,fr}''$ . The latter equation also applies to the present case because the unbound membrane segment (or mother membrane) is characterized by the same fluid-elastic parameters m and  $\kappa$  as in the case of the uniform membrane.

Likewise, for the final spreading process close to the completely engulfed state C, *i.e.*, for small deviations  $\delta \phi \equiv \phi - \pi$ , the free energy landscape for the unbound membrane segment behaves as  $E_{\rm un}(\phi) \approx E_{\rm un}(\pi) + \frac{1}{2} E_{\rm un,ce}'' \delta \phi^2$  with

$$E_{\rm un,ce}'' \equiv \left. \frac{d^2 E(\phi)}{d \phi^2} \right|_{\phi=\pi} = 4\pi \kappa R_{\rm pa} [M_{\rm ms}' - m] \left[ \pm 2 + R_{\rm pa} (3m - M_{\rm ms}') \right].$$
(S28)

which implies the gradient

$$\frac{d E_{\rm un}(\phi)}{d \phi} \approx 4\pi \kappa R_{\rm pa}(M'_{\rm ms} - m) \left[\pm 2 + R_{\rm pa}(3m - M'_{\rm ms})\right] \delta\phi \quad \text{for small } \delta\phi.$$
(S29)

Inspection of Eqs. S27 and S29 shows that the gradient  $d E_{\rm un}(\phi)/d \phi$  is proportional to  $R_{\rm pa}(M_{\rm ms}-m)$  for small  $\phi$  and to  $R_{\rm pa}(M'_{\rm ms}-m)$  for small  $\delta\phi = \phi - \pi$ . These dependencies have two implications. First, the gradient  $d E_{\rm un}(\phi)/d \phi$  vanishes for small  $\phi$  and  $M_{\rm ms} = m$ 

as well as for small  $\delta\phi$  and  $M'_{\rm ms} = m$  as expected. Second, this gradient becomes small if both the segment curvatures  $M_{\rm ms}$  and  $M'_{\rm ms}$  as well as the spontaneous curvature m are small compared to the inverse particle radius  $1/R_{\rm pa}$ . The latter property motivates a systematic expansion of the free energy  $E_{\rm un}$  in powers of the size ratio  $\epsilon \equiv R_{\rm pa}/R_{\rm ve}$  with  $R_{\rm ve} = \sqrt{A/4\pi}$ as before. Such an expansion shows (i) that  $M'_{\rm ms} \approx M_{\rm ms}$  to leading order in  $\epsilon$  and (ii) that the free energy gradient  $d E_{\rm un}/d\phi$  of the unbound membrane segment behaves as

$$\frac{d E_{\rm un}(\phi)}{d \phi} = \pm 8\pi\kappa R_{\rm pa}(M_{\rm ms} - m)\sin(\phi)\cos(\phi) + \mathcal{O}(\epsilon^2).$$
(S30)

For small values of  $\phi$  and  $\delta \phi = \phi - \pi$ , this expression becomes identical, to first order in  $\epsilon = R_{\rm pa}/R_{\rm ve}$ , with Eq. S27 and Eq. S29, respectively. This asymptotic behavior has been confirmed by numerical minimization of the total free energy. Therefore, in the limit of small size ratios  $R_{\rm pa}/R_{\rm ve}$ , the gradient  $dE_{\rm un}(\phi)/d\phi$  is proportional to  $R_{\rm pa}(M_{\rm ms} - m) \approx R_{\rm pa}(M_{\rm ms}' - m)$  for all values of  $\phi$ . As a consequence, this gradient can be neglected if both the segment curvatures  $M_{\rm ms}$  and  $M_{\rm ms}'$  as well as the spontaneous curvature m are much smaller than the inverse particle radius  $1/R_{\rm pa}$ . In the latter case, the gradient of the free energy landscape is determined by the bound membrane segment alone and behaves as

$$\frac{d E(\phi)}{d \phi} \approx \frac{d E_{\rm bo}(\phi)}{d \phi} = \left[-2\pi |W| R_{\rm pa}^2 + 4\pi \kappa_{\rm bo} (1 \pm m_{\rm bo} R_{\rm pa})^2\right] \sin(\phi) \tag{S31}$$

as follows from Eq. S26. When we insert this expression for  $d E(\phi)/d \phi$  into the equation of motion as given by Eq. S24 (or Eq. 17 in the main text), the factors proportional to  $\sin(\phi)$ cancel and we obtain the simplified equation of motion

$$\frac{d\phi}{dt} = \frac{|W|R_{\rm pa}^2 - 2\kappa(1 \pm mR_{\rm pa})^2}{\eta_{\rm eff} R_{\rm pa}^3}$$
(S32)

which implies that the angular velocity  $d\phi/dt$  is constant and that the engulament time  $t_{\mathcal{FC}}$ 

follows from

$$\pi = \frac{|W|R_{\rm pa}^2 - 2\kappa (1 \pm mR_{\rm pa})^2}{\eta_{\rm eff} R_{\rm pa}^3} t_{\mathcal{FC}} \,. \tag{S33}$$

For the plus sign corresponding to endocytosis, Eq. S33 is equivalent to Eq. 18 in the main text.

#### References

- Deuling, H.; Helfrich, W. The Curvature Elasticity of Fluid Membranes: A Catalogue of Vesicle Shapes. J. Physique 1976, 37, 1335–1345.
- (2) Seifert, U.; Berndl, K.; Lipowsky, R. Shape Transformations of Vesicles: Phase Diagram for Spontaneous Curvature and Bilayer Coupling Model. *Phys. Rev. A* 1991, 44, 1182– 1202.
- (3) Lipowsky, R. Coupling of Bending and Stretching Deformations in Vesicle Membranes. Adv. Colloid Interface Sci. 2014, 208, 14–24.
- (4) Berndl, K.; Käs, J.; Lipowsky, R.; Sackmann, E.; Seifert, U. Shape Transformations of Giant Vesicles: Extreme Sensitivity to Bilayer Asymmetry. *Europhys. Lett.* 1990, 13, 659–664.
- (5) Lipowsky, R. The Conformation of Membranes. *Nature* **1991**, *349*, 475–481.
- (6) Döberreiner, H.-G.; Evans, E.; Kraus, M.; Seifert, U.; Wortis, M. Mapping Vesicle Shapes into the Phase Diagram: A Comparison of Experiment and Theory. *Phys. Rev.* E 1997, 55, 4458 – 4474.
- (7) Deserno, M. Elastic Deformation of a Fluid Membrane upon Colloid Binding. *Phys. Rev. E* 2004, 69, 031903.
- (8) Zhang, S.; Li, J.; Lykotrafitis, G.; Bao, G.; Suresh, S. Size-Dependent Endocytosis of Nanoparticles. Adv. Mater. 2009, 21, 419 – 424.

- (9) Tinevez, J.-Y.; Schulze, U.; Salbreux, G.; Roensch, J.; Joanny, J.-F.; Paluch, E. Role of Cortical Tension in Bleb Growth. Proc. Nat. Acad. Sci. USA 2009, 106, 18581–18586.
- (10) Fischer-Friedrich, E.; Hyman, A. A.; Jülicher, F.; Muller, D. J.; Helenius, J. Quantification of Surface Tension and Internal Pressure Generated by Single Mitotic Cells. *Scientific Rep.* 2014, 4, 6213.
- (11) Jin, A. J.; Prasad, K.; Smith, P. D.; Lafer, E. M.; Nossal, R. Measuring the Elasticity of Clathrin-Coated Vesicles via Atomic Force Microscopy. Biophys. J. 2006, 90, 3333 3344.
- (12) Chithrani, B. D.; Ghazani, A. A.; Chan, W. C. W. Determining the Size and Shape Dependence of Gold Nanoparticle Uptake into Mammalian Cells. *Nano Lett.* 2006, 6, 662 – 668.
- (13) Chithrani, B. D.; Chan, W. C. W. Elucidating the Mechanism of Cellular Uptake and Removal of Protein-Coated Gold Nanoparticles of Different Sizes and Shapes. Nano Lett. 2007, 7, 1542 – 1550.